

Moleculaire diagnostiek Deel 2

Marjan vd Werken

Karlijn Stouten

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Programma

Opfrissen deel 1 van vorig jaar: Rob

Introductie verworven mutaties: Rob

Inleiding 'integrale diagnostiek' en myeloproliferatieve aandoeningen: Rob

Achtergrond complexere DNA-technieken: Marjan

Uitleg over nieuwe DNA-ruimten: Marjan

Uitleg en resultaten van onderzoek Karlijn: Karlijn

Casuïstiek: Marjan en Rob



Moleculaire diagnostiek

Met laboratoriumtesten de genetische code (DNA) onderzoeken.

=

een van de speerpunten van Result Laboratorium

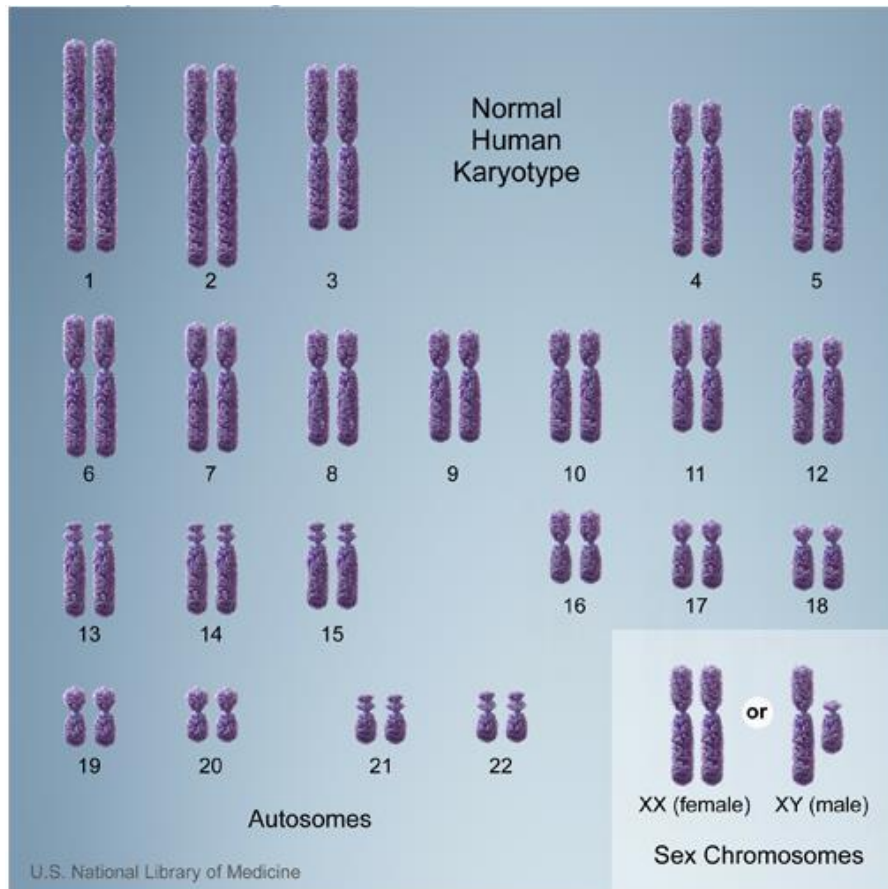


DNA: de genetische code **“een receptenboek”**

- DNA-code bestaat uit (maar) vier verschillende letters:
A, T, G en C
- Menselijke code is drie miljard (3.000.000.000) letters lang.
- Alle DNA van een cel uitgerold: snoer van 1 - 1,5 meter.
- Klein gedeelte (1%) van de DNA-code zijn genen.
- Aantal genen van een mens: circa 22.000.
- Een gen is het recept voor een eiwit.
- Ieder mens heeft zijn eigen, unieke, DNA-code.
- Tussen mensen verschilt DNA-code van genen 0,1%.



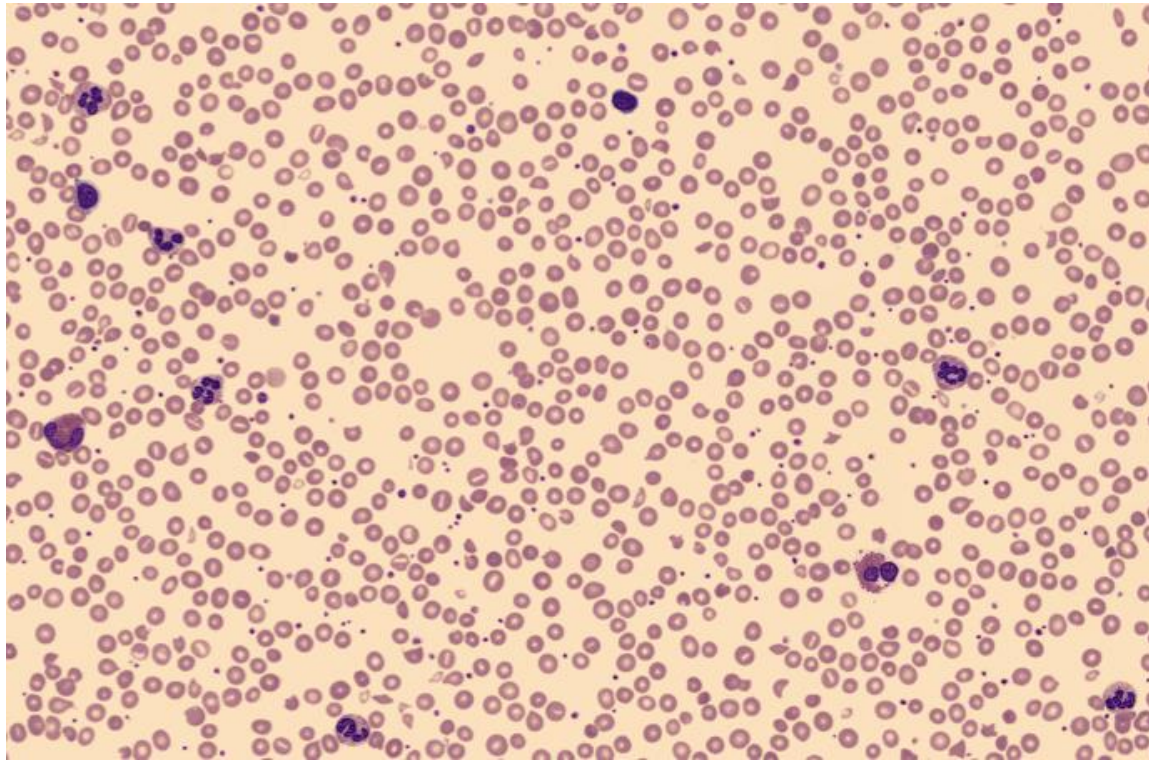
DNA: niet een aaneengesloten snoer, maar verdeeld over chromosomen



Van elk chromosoom twee exemplaren:
een van je vader en een van je moeder.



De 46 chromosomen zitten in de celkern.

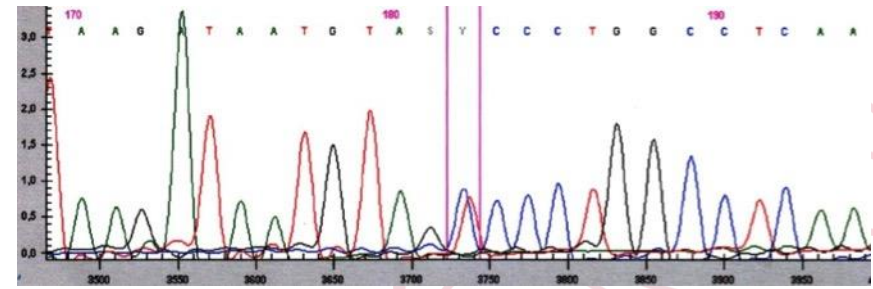
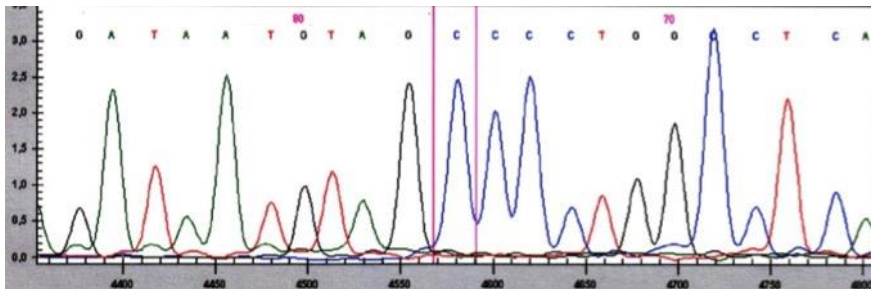


Blijft je genetisch code hetzelfde?

De DNA-code erf je van je ouders,
maar tijdens je leven ontstaan in sommige cellen veranderingen in
de genetische code.

Uitdaging:

Hoe toon je die verworven mutaties aan?



Moleculaire diagnostiek wordt een steeds belangrijker onderdeel van ‘integrale diagnostiek’.

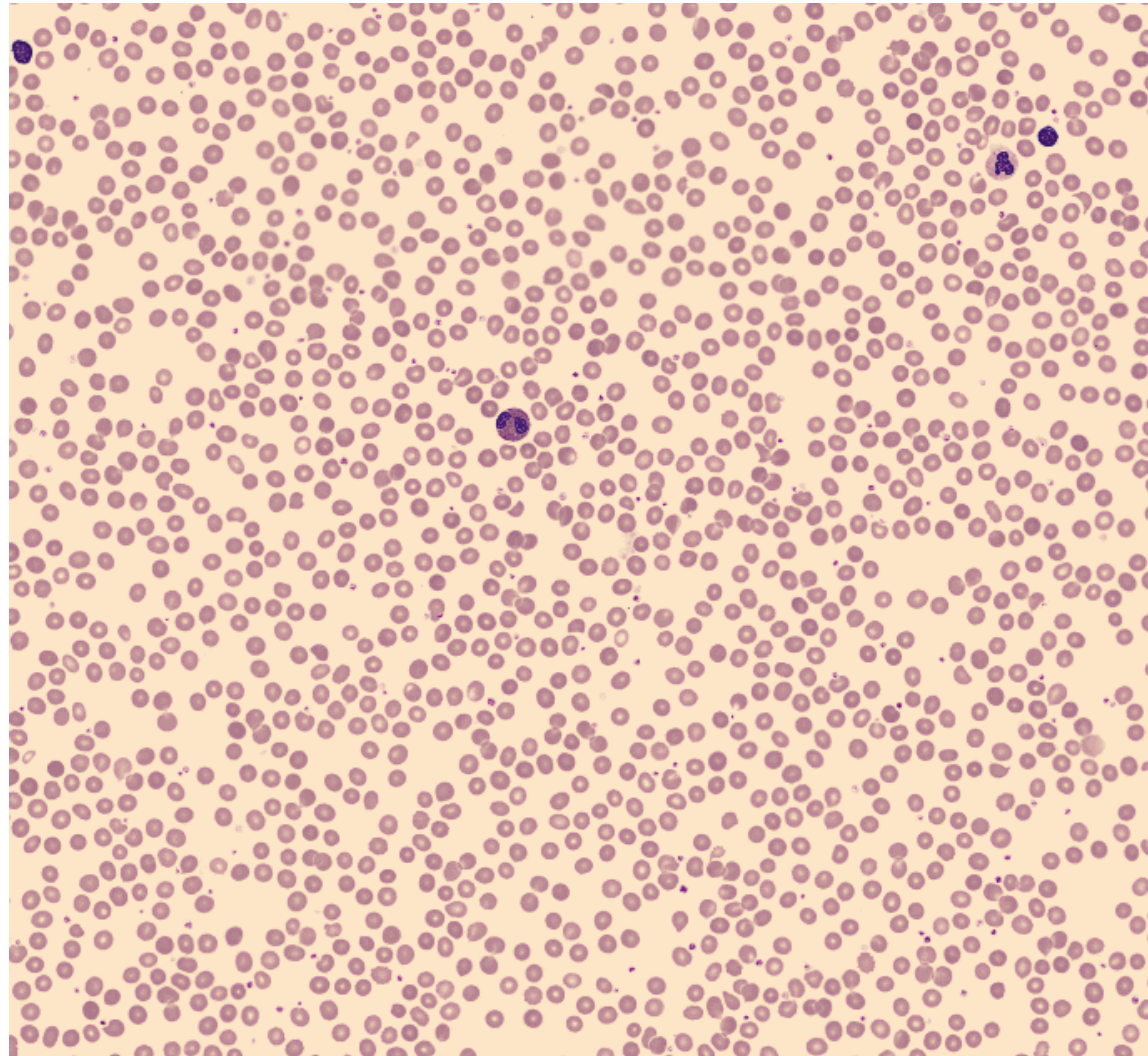
Bijvoorbeeld: hemato-oncologie.

- lichamelijk onderzoek
- medische voorgeschiedenis
- routine laboratoriumtesten
 - morfologie
 - flowcytometrie
- moleculaire diagnostiek



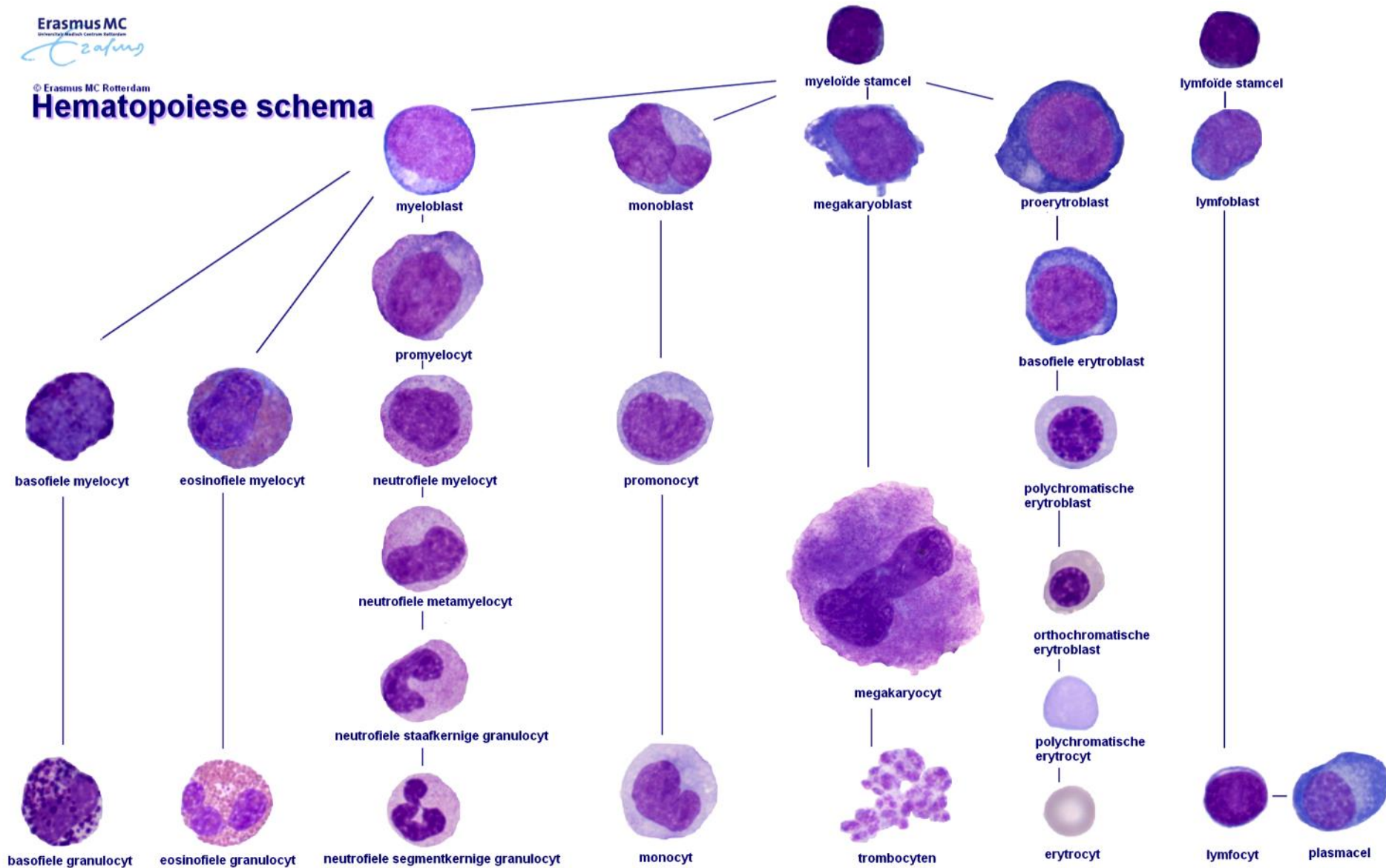
Normale verhouding cellen bloed.

Rode bloedcellen ($4,2-6,2 \times 10^{12}/L$)
Witte bloedcellen ($4,3-10 \times 10^9/L$)
Bloedplaatjes ($150-400 \times 10^9/L$)



Hematopoiese

Hematopoiese schema

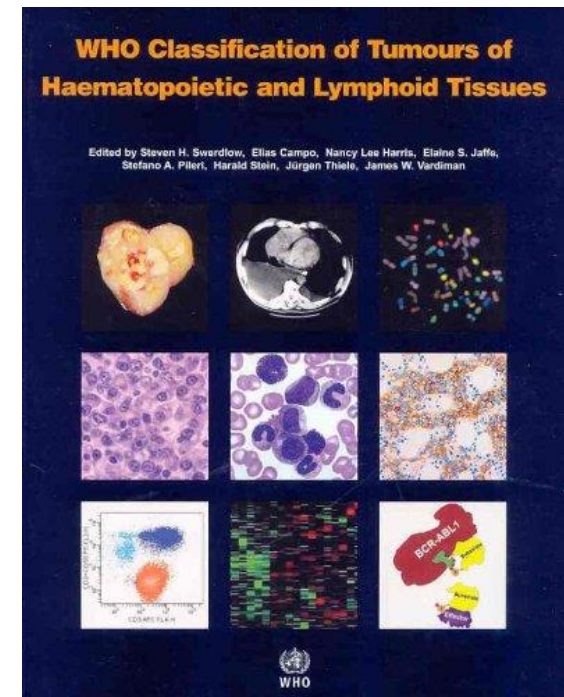


Myeloproliferatieve ziekten

MPN indeling volgens de 3^{de} WHO editie (2008)

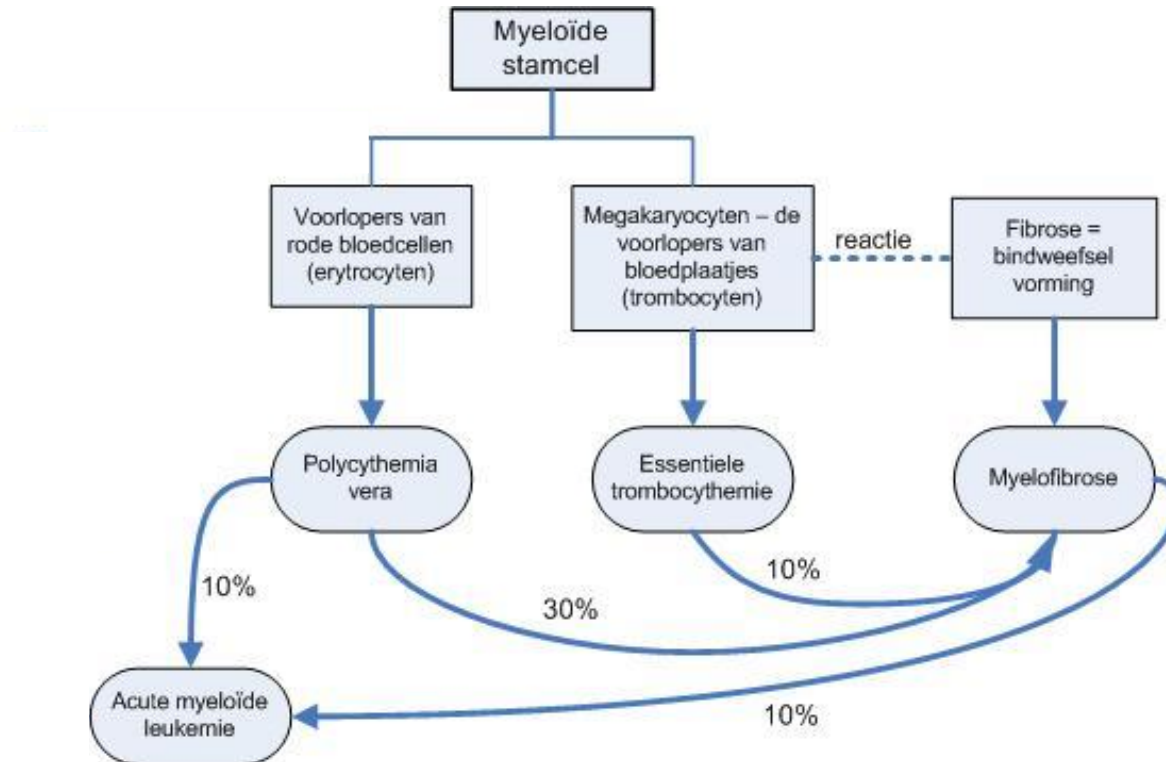
Tabel 1. Myeloproliferatieve neoplasmata (MPN).

chronische myeloïde leukemie, BCR-ABL1-positief (CML)	
chronische neutrofiele leukemie	
polycythemia vera (PV)	'Klassieke MPN'
primaire myelofibrose (MF)	
essentiële trombocytose (ET)	
chronische eosinofiele leukemie, niet nader gespecificeerd (CEL NOS)	
mastocytose	
myeloproliferatief neoplasma, niet classificeerbaar	



Mastocytose: Aparte categorie in nieuwe 4^{de} WHO editie 2016.

Woekering van één (of meerdere) cellijnen.



Klinische problemen: verhoogd risico op veneuze en arteriële trombotische complicaties, en op bloedingen.

Prognose

PV

In het algemeen een gemiddelde overleving na diagnose van 10 - 20 jaar.

ET

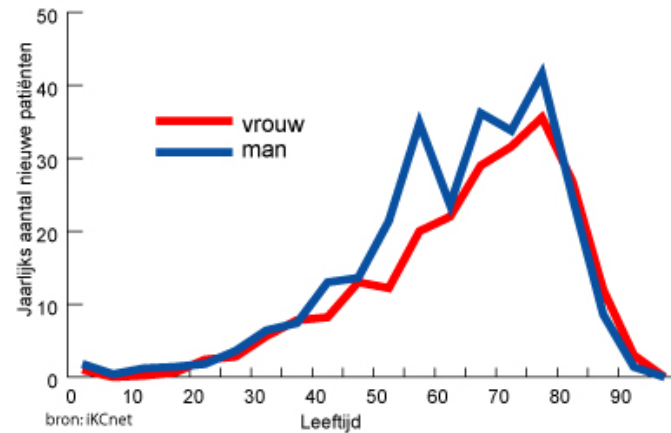
Bij een goede behandeling, zal de levensverwachting van patiënten met ET niet sterk afwijken van die van mensen zonder ET.

MF

Patiënten kunnen de ziekte 10 jaar of langer overleven, vooral als de diagnose vroeg gesteld wordt. Zeer ernstige gevallen kunnen binnen 3 tot 6 jaar fataal zijn.



Zeldzaam?

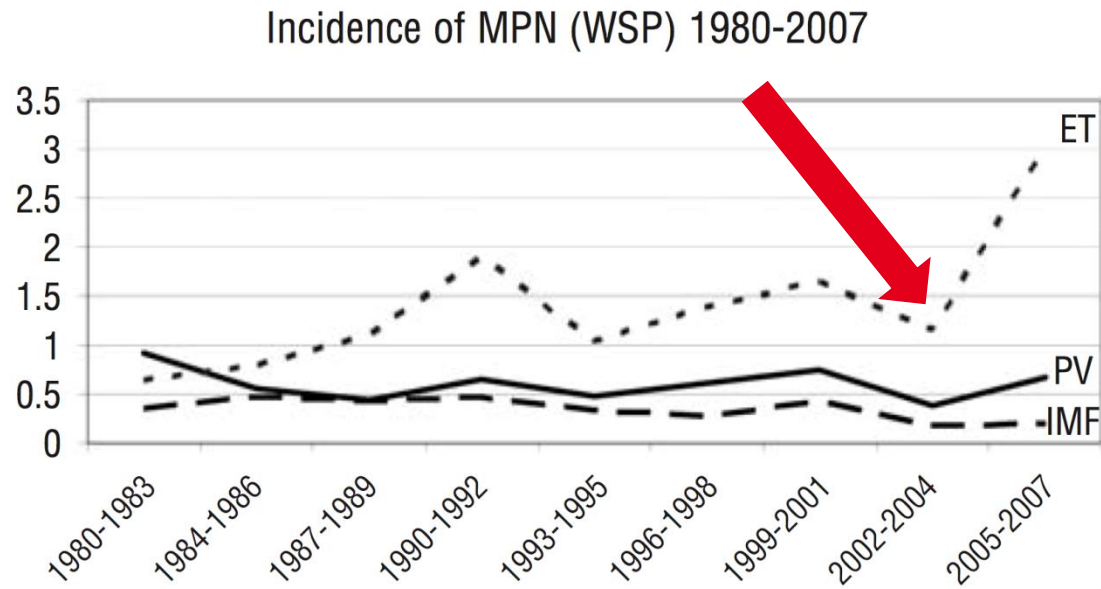


- Voornamelijk >50 jaar, maar van alle leeftijden
- Incidentie: 6-10/100.000/jaar
- Jaarlijks 1.000 nieuwe patiënten in Nederland

Maar...



Incidentie neemt toe



F. Girodon et al. Haematologica 2009; 94(6)

2005
betere laboratoriumdiagnostiek:
moleculaire diagnostiek.



Onderscheid primair/secundair kan lastig zijn...

Tabel 8.22 Oorzaken van trombocytose.

primair:

- essentiële trombocytose*
- andere myeloproliferatieve ziekten*

secundair:

- postoperatief
- na grote bloedingen
- (chronische) infecties
- Fe-gebrek
- niet-hematologische maligniteiten

Tabel 3. Oorzaken van congenitale of verworven secundaire erythrocytose.

oorzaken congenitale secundaire erythrocytose

- erytropoëtineceptordefect
- hemoglobinevariant met verhoogde zuurstofaffiniteit
- bisfosfoglyceraatmutasedeficiëntie
- verstoorde regulatie van de zuurstofhomeostase door 'hypoxia inducible factor' (HIF); mutaties in von hippel-lindau-tumorsuppressor (chuvash-erythrocytose), prolylhydroxylase-2 en HIF2- α

oorzaken van verworven secundaire erythrocytose

- centrale hypoxie (bijvoorbeeld door chronische longziekte, roken, slaapapneusyndroom, koolmonoxidevergiftiging, verblijf op grote hoogte)
- lokale, renale hypoxie (bijvoorbeeld nierarteriestenose, cyste nieren, postniertransplantatie)
- pathologische erytropoëtineproductie (bijvoorbeeld door cerebellair hemangioblastoom, meningioom, niercelcarcinoom, hepatocellulair carcinoom)
- exogene toediening van erytropoëtine

Moleculaire diagnostiek

2005: JAK2 V617F mutatie (95% PV, 60% ET, 50% MF)

2006: MPL W515L/K (5-10% ET en MF)

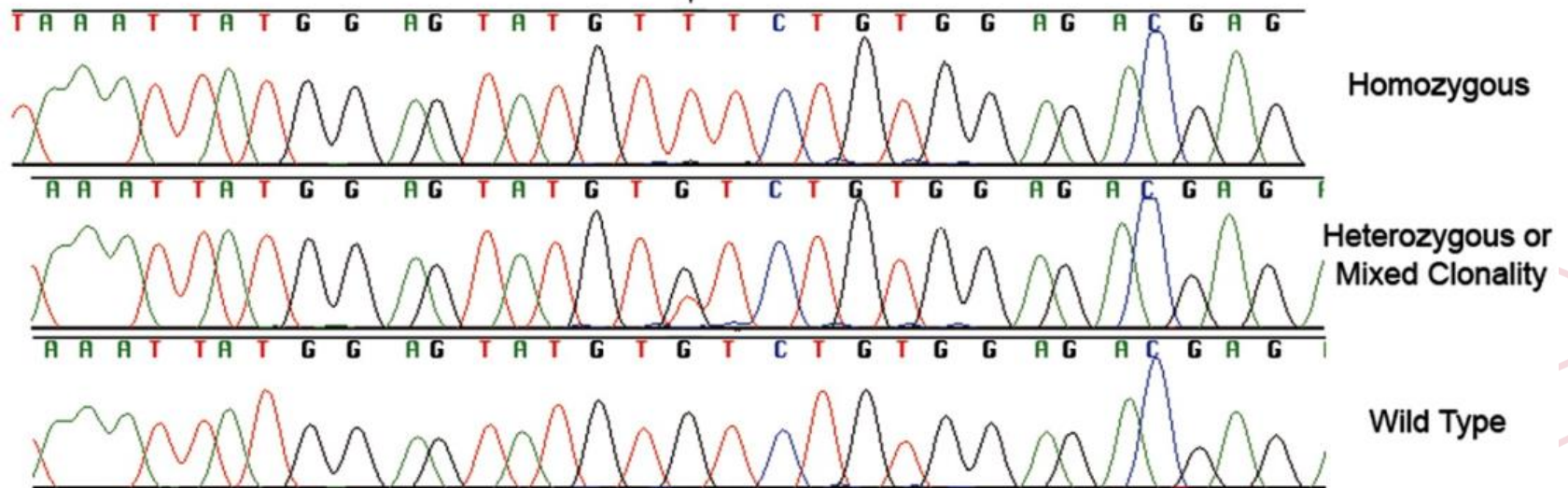
2007: JAK2 exon 12 (5% PV)

2013: CALR exon 9 (25% ET en MF)



JAK2 V617F

JAK2 V617F Mutation



JAK2 exon 12 mutaties

SUBSTITUTIONS

wildtype

F533IK539L

F537IK539L

H538QK539L

H538DK539LI540S

K539L

K539LL545V

I540T

D544G

L545S

F547L

F547V

(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMTQMVFHLIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVTHLIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVQLIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFDLSRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVHLIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVHLIRNEDVIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKTRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRNEGLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRNEDSIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRNEDLIL
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRNEDLIV

DELETIONS

wildtype

F537-K539delinsK

F537-K539del

F537-K539delinsL

H538del

H538-K539del

H538-K539delinsF

H538-K539delinsI

H538-K539delinsL

I540-N542delinsS

I540-N542delinsK

I540-E543delinsKK

I540-E543delinsMK

I540-D544delinsMK

I540S, R541-E543delinsK

R541-E543delinsK

R541-D544del

N542-E543del

N542-D544delinsN

E543-D544del

D544-L545del

(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVLIIRNEDLIF
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(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKSEDLIF
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(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKMKDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKMKLIF
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(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRDIF
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(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRNLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVMVFKIRNEIF

DUPLICATIONS

wildtype

V536-I546dup11

V536-F547dup12

V536F, F37-I546dup10

F537-F547dup11

F537-I546dup10, F547L

F547L, I540-F547dup8

(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIVFHKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIVFHKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF
(D) KSNLLVFRITNGVSDVPTSPITLQRPTHMNMVVFHKIRNEDLIFVFKIRNEDLIF

MPL mutaties

W515K

W515L

V501A

S505C

A506T

V507I

G509C

L510P

R514K

W515R

R519T

S204P

Y591N

S505N



CALR exon 9 mutaties

Type 1	TRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 2	NCRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 3	QTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 4	RRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 5	GQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 6	RRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 7	RRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 8	RRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 9	RQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 10	MCRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 11	DQRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 12	RRRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 13	QRRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 14	RRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 15	RRRERTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 16	QRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 17	RRQWTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 18	RMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 19	RQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 20	GRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 21	AFKTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 22	NAKRRRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 23	CVRRRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 24	RRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 25	RQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 26	NAKRRRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 27	CFAKRRRRQTRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 28	RRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 29	PPLCLRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 30	DHPCRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 31	GNCRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 32	CRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 33	CRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 34	TCCRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 35	ICRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-
Type 36	CRRMMRTKMRMRMRRTTRKMRKMSPARPRTSCREACLQGWTEA-

Table 4. WHO criteria for PV

WHO PV criteria

Major criteria

1. Hemoglobin >16.5 g/dL in men 10,2 mmol/L

Hemoglobin >16.0 g/dL in women 9,9 mmol/L

or,

Hematocrit >49% in men

Hematocrit >48% in women

or,

increased red cell mass (RCM)*

2. BM biopsy showing hypercellularity for age with trilineage growth (panmyelosis) including prominent erythroid, granulocytic, and megakaryocytic proliferation with pleomorphic, mature megakaryocytes (differences in size)

3. Presence of *JAK2V617F* or *JAK2* exon 12 mutation

Minor criterion

Subnormal serum erythropoietin level

Diagnosis of PV requires meeting either all 3 major criteria, or the first 2 major criteria and the minor criterion†

*More than 25% above mean normal predicted value.

†Criterion number 2 (BM biopsy) may not be required in cases with sustained absolute erythrocytosis: hemoglobin levels >18.5 g/dL in men (hematocrit, 55.5%) or >16.5 g/dL in women (hematocrit, 49.5%) if major criterion 3 and the minor criterion are present. However, initial myelofibrosis (present in up to 20% of patients) can only be detected by performing a BM biopsy; this finding may predict a more rapid progression to overt myelofibrosis (post-PV MF).



Table 5. WHO criteria for ET

WHO ET criteria

Major criteria

1. Platelet count $\geq 450 \times 10^9/L$
2. BM biopsy showing proliferation mainly of the megakaryocyte lineage with increased numbers of enlarged, mature megakaryocytes with hyperlobulated nuclei. No significant increase or left shift in neutrophil granulopoiesis or erythropoiesis and very rarely minor (grade 1) increase in reticulin fibers
3. Not meeting WHO criteria for *BCR-ABL1*⁺ CML, PV, PMF, myelodysplastic syndromes, or other myeloid neoplasms
4. Presence of *JAK2*, *CALR*, or *MPL* mutation

Minor criterion

Presence of a clonal marker or absence of evidence for reactive thrombocytosis

Diagnosis of ET requires meeting all 4 major criteria or the first 3 major criteria and the minor criterion



Table 6. WHO criteria for prePMF

WHO prePMF criteria

Major criteria

1. Megakaryocytic proliferation and atypia, without reticulin fibrosis >grade 1*, accompanied by increased age-adjusted BM cellularity, granulocytic proliferation, and often decreased erythropoiesis
2. Not meeting the WHO criteria for *BCR-ABL1*⁺ CML, PV, ET, myelodysplastic syndromes, or other myeloid neoplasms
3. Presence of *JAK2*, *CALR*, or *MPL* mutation or in the absence of these mutations, presence of another clonal marker,† or absence of minor reactive BM reticulin fibrosis‡

Minor criteria

Presence of at least 1 of the following, confirmed in 2 consecutive determinations:

- a. Anemia not attributed to a comorbid condition
- b. Leukocytosis $\geq 11 \times 10^9/L$
- c. Palpable splenomegaly
- d. LDH increased to above upper normal limit of institutional reference range

Diagnosis of prePMF requires meeting all 3 major criteria, and at least 1 minor criterion

*See Table 8.

†In the absence of any of the 3 major clonal mutations, the search for the most frequent accompanying mutations (eg, *ASXL1*, *EZH2*, *TET2*, *IDH1/IDH2*, *SRSF2*, *SF3B1*) are of help in determining the clonal nature of the disease.

‡Minor (grade 1) reticulin fibrosis secondary to infection, autoimmune disorder or other chronic inflammatory conditions, hairy cell leukemia or other lymphoid neoplasm, metastatic malignancy, or toxic (chronic) myelopathies.

Table 7. WHO criteria for overt PMF

WHO overt PMF criteria

Major criteria

1. Presence of megakaryocytic proliferation and atypia, accompanied by either reticulin and/or collagen fibrosis grades 2 or 3*
2. Not meeting WHO criteria for ET, PV, *BCR-ABL1*⁺ CML, myelodysplastic syndromes, or other myeloid neoplasms
3. Presence of *JAK2*, *CALR*, or *MPL* mutation or in the absence of these mutations, presence of another clonal marker,† or absence of reactive myelofibrosis‡

Minor criteria

Presence of at least 1 of the following, confirmed in 2 consecutive determinations:

- a. Anemia not attributed to a comorbid condition
- b. Leukocytosis $\geq 11 \times 10^9/L$
- c. Palpable splenomegaly
- d. LDH increased to above upper normal limit of institutional reference range
- e. Leukoerythroblastosis

Diagnosis of overt PMF requires meeting all 3 major criteria, and at least 1 minor criterion

*See Table 8.

†In the absence of any of the 3 major clonal mutations, the search for the most frequent accompanying mutations (eg, *ASXL1*, *EZH2*, *TET2*, *IDH1/IDH2*, *SRSF2*, *SF3B1*) are of help in determining the clonal nature of the disease.

‡BM fibrosis secondary to infection, autoimmune disorder, or other chronic inflammatory conditions, hairy cell leukemia or other lymphoid neoplasm, metastatic malignancy, or toxic (chronic) myelopathies.

Table 7. WHO criteria for overt PMF

WHO overt PMF criteria

Major criteria

1. Presence of megakaryocytic proliferation and atypia, accompanied by either reticulin and/or collagen fibrosis grades 2 or 3*
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3. Presence of *JAK2*, *CALR*, or *MPL* mutation or in the absence of these mutations, presence of another clonal marker,† or absence of reactive myelofibrosis‡

Minor criteria

Presence of at least 1 of the following, confirmed in 2 consecutive determinations:

- a. Anemia not attributed to a comorbid condition
- b. Leukocytosis $\geq 11 \times 10^9/L$
- c. Palpable splenomegaly
- d. LDH increased to above upper normal limit of institutional reference range
- e. Leukoerythroblastosis

Diagnosis of overt PMF requires meeting all 3 major criteria, and at least 1 minor criterion

*See Table 8.

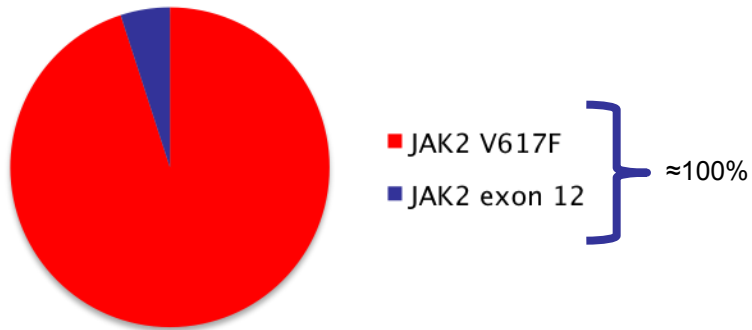
†In the absence of any of the 3 major clonal mutations, the search for the most frequent accompanying mutations (eg, *ASXL1*, *EZH2*, *TET2*, *IDH1/IDH2*, *SRSF2*, *SF3B1*) are of help in determining the clonal nature of the disease.

‡BM fibrosis secondary to infection, autoimmune disorder, or other chronic inflammatory conditions, hairy cell leukemia or other lymphoid neoplasm, metastatic malignancy, or toxic (chronic) myelopathies.

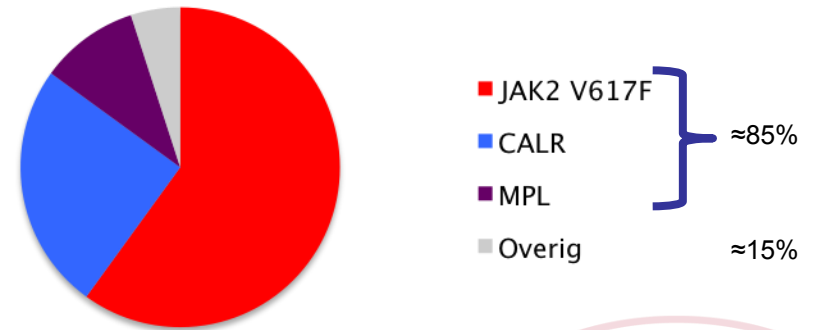


Moleculaire diagnostiek

PV



ET / MF

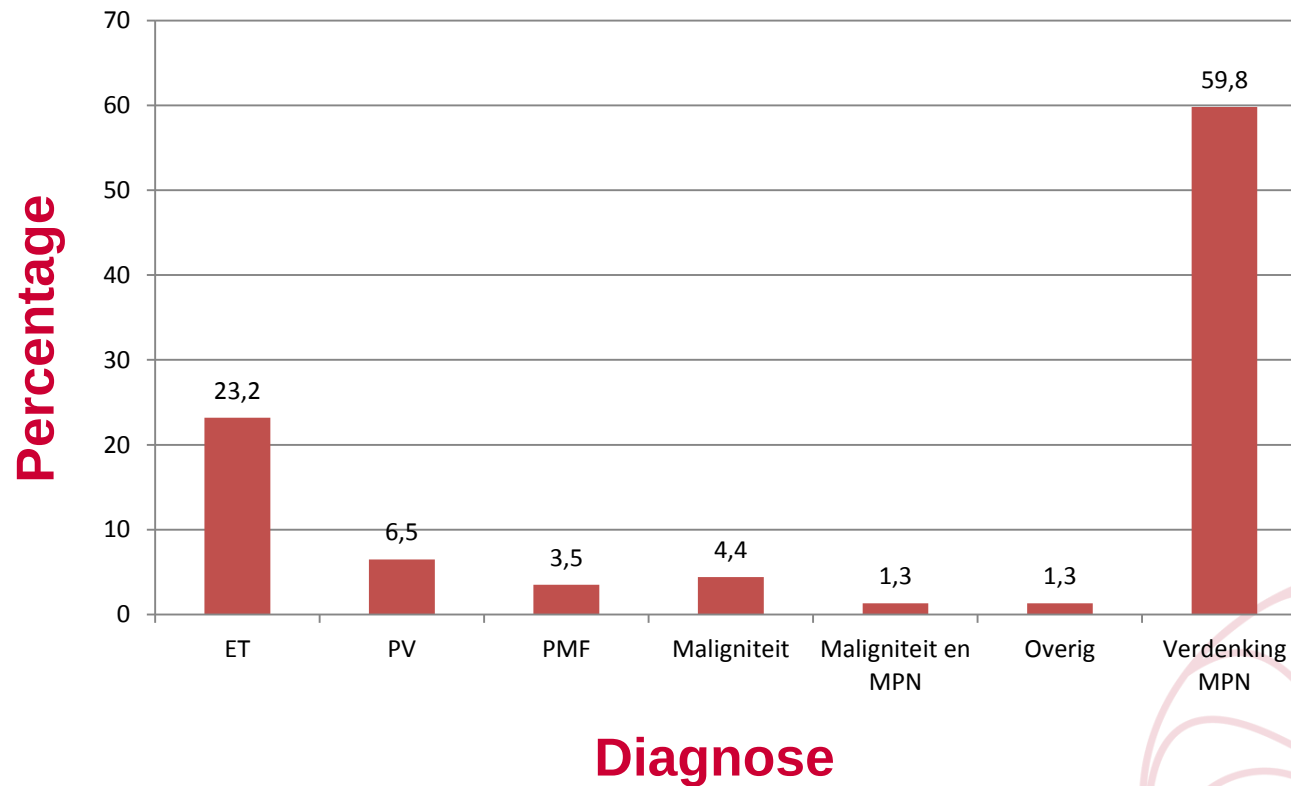


Database Result

- DNA van alle patiënten waarvoor een JAK2 V617F bepaling is aangevraagd
- 981 patiënten sinds maart 2012
- Gevonden mutatie en gebruikte techniek worden geregistreerd
- 850 patiënten (86.6%) hebben bekende diagnose

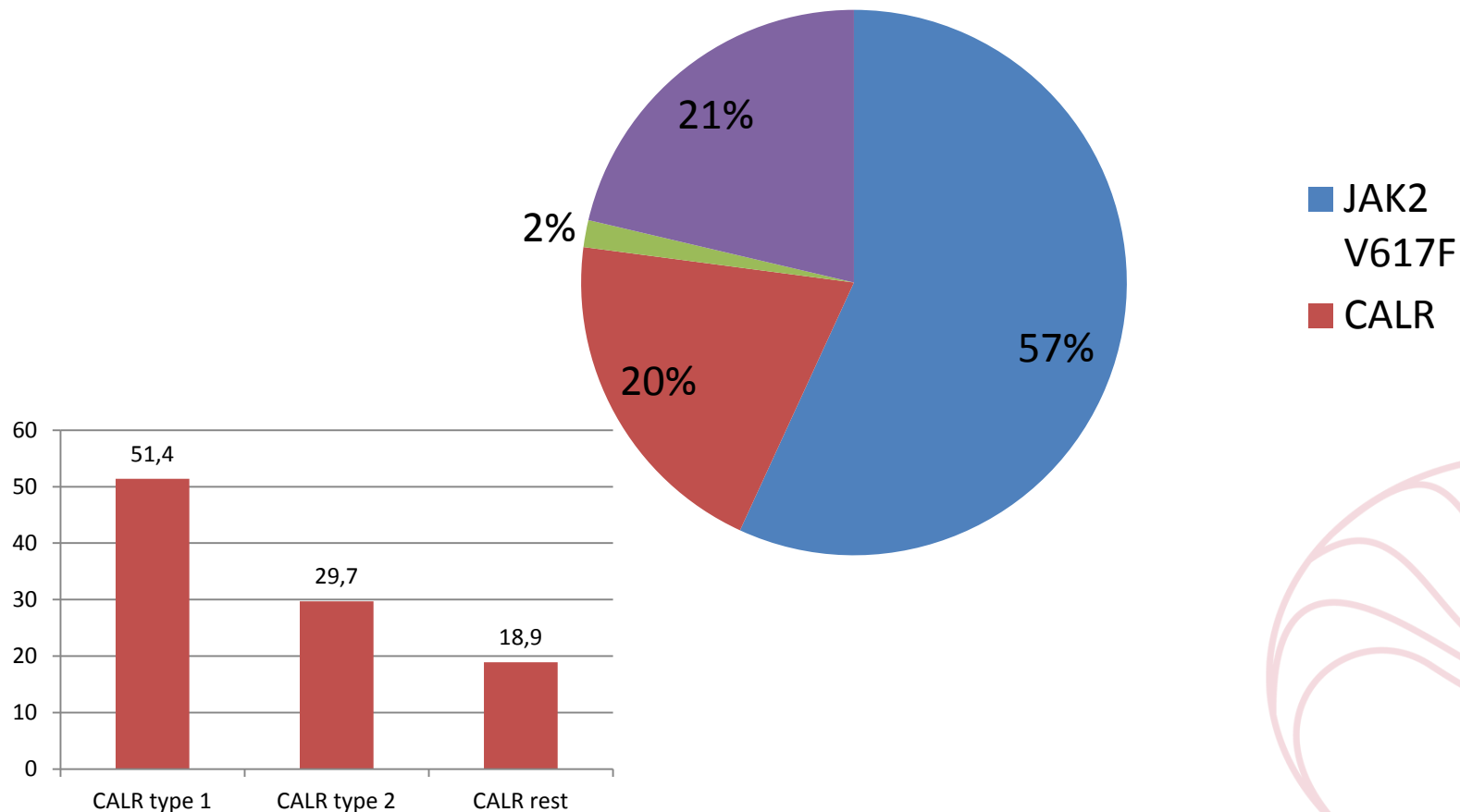


Diagnoses



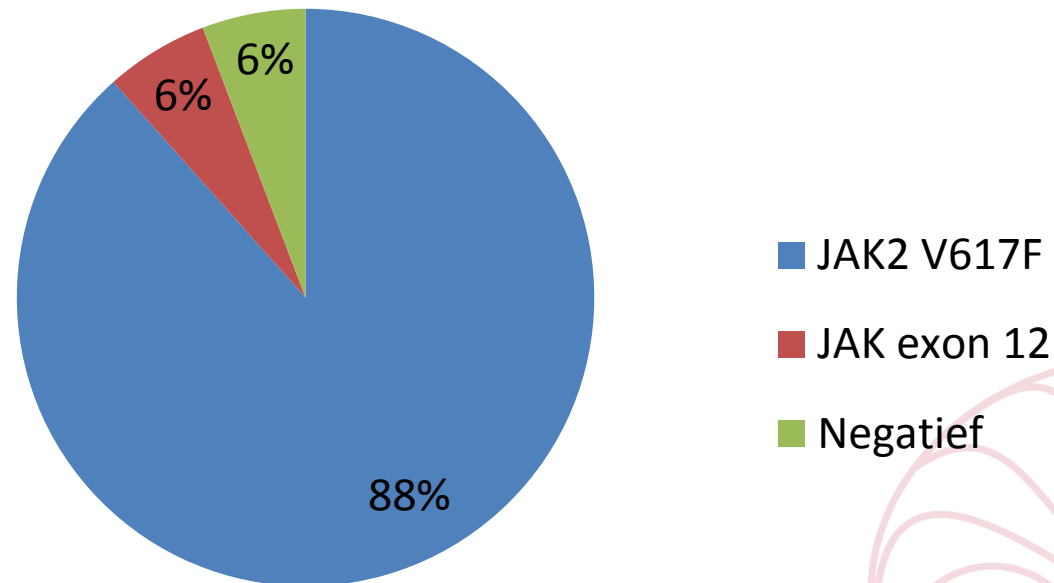
Mutaties ET

- 197 patiënten met ET → 182 volledig getest



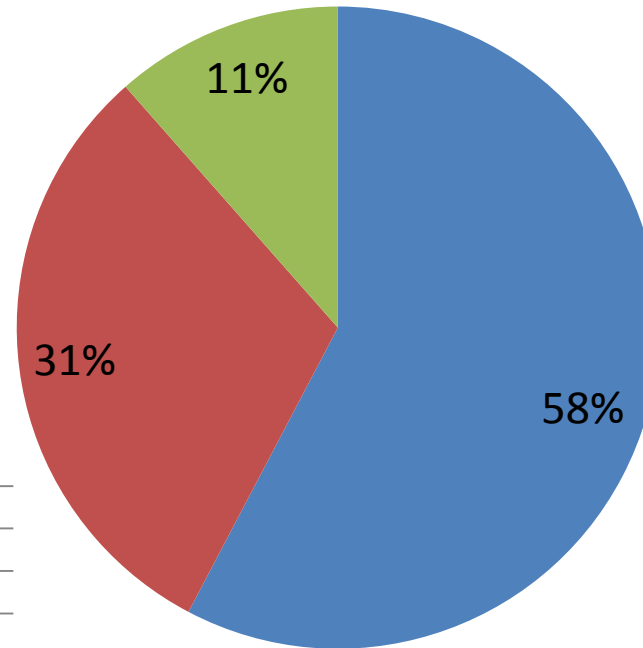
Mutaties PV

- 55 patiënten met PV → 52 volledig getest

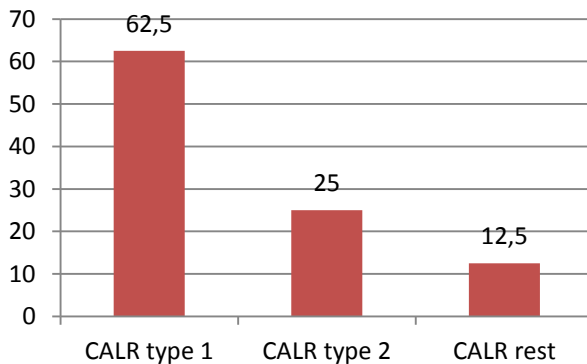


Mutaties PMF

- 30 patiënten met PMF → 26 volledig getest



■ JAK2 V617F
■ CALR
■ MPL



- JAK2 V617F
- JAK2 exon 12
- Calreticuline (CALR)
- MPL mutaties



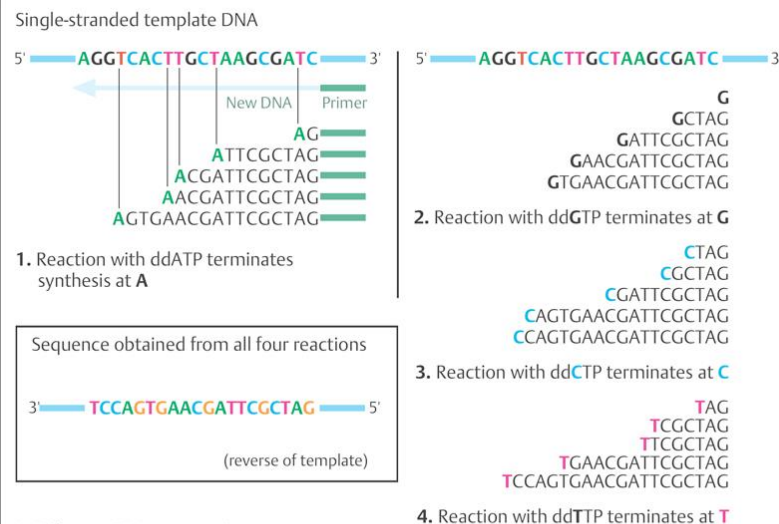
Technieken voor MPN onderzoek

- Sequentie analyse
- MLPA
- Real Time
- PCR

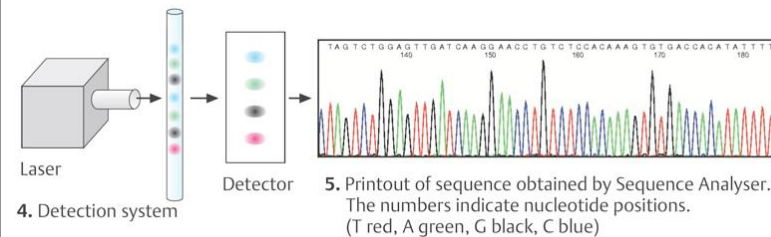
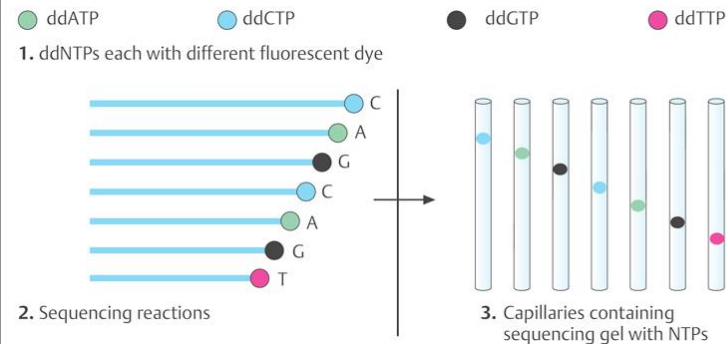


Sequentie analyse





A. Dideoxy DNA sequencing



B. Automated DNA sequencing

Voordelen

Toont puntmutaties en kleine deleties of inserties goed aan

Nadelen

Toont geen grote deleties of inserties aan

Lage gevoeligheid (20%)





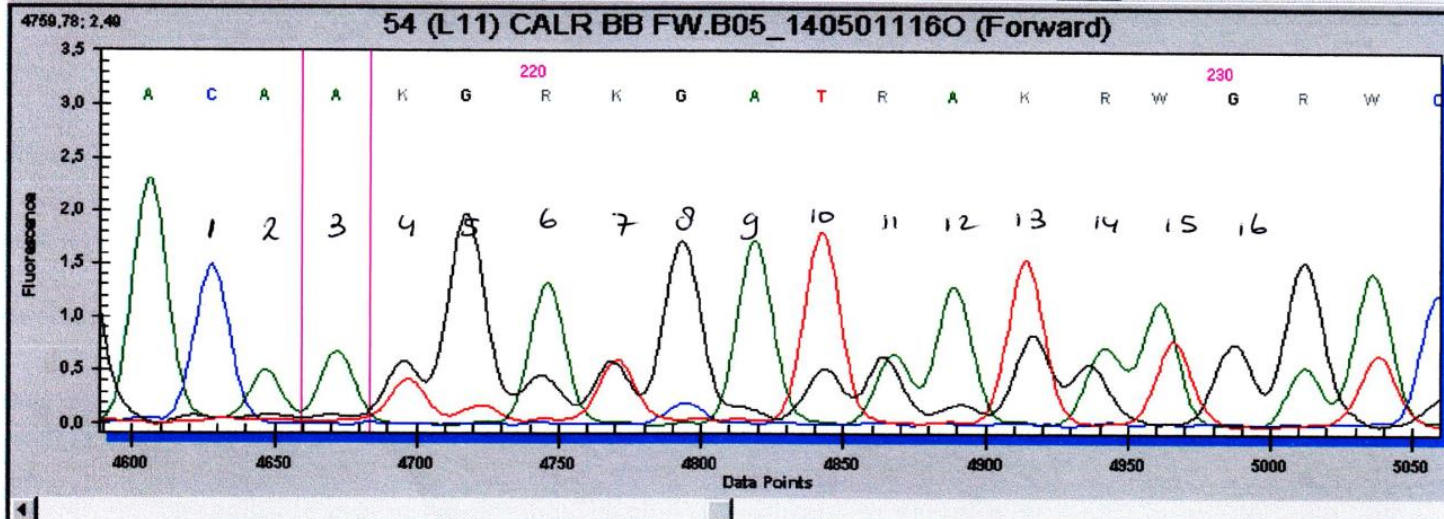
CALR mutaties



Type 1	c.1092_1143del	→ 52 bp deletie
Type 2	c.1154_1155insTTGTC	→ 5bp insertie TTGTC
Type 3	c.1095_1140del	→ 46 bp deletie
Type 4	c.1102_1135del	→ 34 bp deletie
Type 5	c.1091_1142del	→ 52 bp deletie
Type 6	c.1094_1139del	→ 46 bp deletie
Type 7	c.1102_1153del	→ 52 bp deletie
Type 8	c.1104_1137del	→ 34 bp deletie
Type 9	c.1140del	→ 1 bp deletie
Type 10	c.1154delinsTGTGTC	→ 1 bp deletie en insertie van TGTGTC
Type 11	[c.1092G>C;1095_1140del]	→ 1092 G wordt C en deletie van 46 bp
Type 12	c.1098_1131del	→ 34 bp deletie
Type 13	c.1100_1134delinsA	→ 35 bp deletie en insertie A
Type 14	c.1101_1134del	→ 34 bp deletie
Type 15	[c.1102_1135del;1145C>G]	→ 34 bp deletie en C wordt G
Type 16	c.1102_1137delinsCA	→ 36 bp deletie en insertie CA
Type 17	[c.1104_1137del;1148A>T]	→ 34 bp deletie en A wordt T



Base Number	310	320	330	340
Codon Number	102 103 104 105 106 107 108 109 110 111 112 113 114	115 116 117 118 119 120 121 122 123 124 125 126 127	128 129 130 131 132 133 134 135 136 137 138 139 140	141 142 143 144 145 146 147 148 149 150 151 152 153
Reference AA Translation	E D K E D D E D K D E D E	E D D E D D E D K D E D E	E D D E D D E D K D E D E	E D D E D D E D K D E D E
CALR BB FW+ CALR AA Rev.txt	CA GAG GAC AAG GAG GAT GAT GAG GAC AAA G-AT GAG GAT G	CA GAG GAC AAG GAG GAT GAT GAG GAC AAA G-AT GAG GAT G	CA GAG GAC AAG GAG GAT GAT GAG GAC AAA G-AT GAG GAT G	CA GAG GAC AAG GAG GAT GAT GAG GAC AAA G-AT GAG GAT G
54 (L11) CALR BB FW.B05_140501	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F
Differences		? ??	? ? ??	?? ? <+? ? ?? ?
Consensus	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F	CA GAG GAC AAK GRK GAT RAK RWG RWC AAR GAMT RAG RWT F
Consensus AA Translation	E D KN * D * * * K DE X * X	E D KN * D * * * K DE X * X	E D KN * D * * * K DE X * X	E D KN * D * * * K DE X * X



L11 = C.1154_1155 ins TTgTC

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
C	A	A	G	G	A	G	G	A	T	G	A	T	G	A	G
C	A	A	T	T	G	T	C	G	G	A	G	G	A	T	G

↓
inserie.

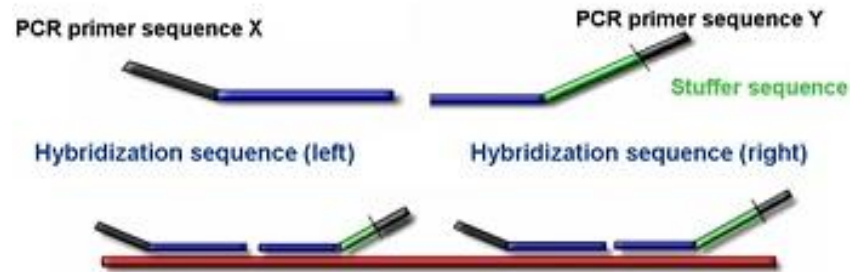
MLPA

Multiplex ligation-dependent probe amplification



Principe MLPA

1. Denaturation and Hybridization



2. Ligation

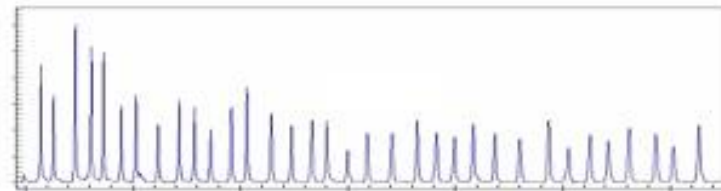


3. PCR with universal primers X and Y

exponential amplification of ligated probes only



4. Fragment analysis





MLPA

Voordelen

Hoge gevoeligheid (tot 0,1%)

Kan goed als screening worden ingezet.

Beperking

Afhankelijkheid van de soort probes die in de mix zitten



Probes in MPN kits

P-520

CALR deletie (type I)
CALR insertie (type II)
MPL W515K
MPL W515L
JAK2 exon 12 (2 mutaties)
JAK2 V617F
KIT D816V



Table 1. SALSA MLPA P520-A1 MPN mix 2

Length (nt)	SALSA MLPA probe	Reference	Chromosomal position	Target	Mutation details
64-70-76-82	Q-fragments: DNA quantity; only visible with less than 100 ng sample DNA				
88-92-96	D-fragments: Low signal of 88 or 96 nt fragment indicates incomplete denaturation				
100	X-fragment: Specific for the X chromosome				
105	Y-fragment: Specific for the Y chromosome				
115	Reference probe S0973-L26704	4p13			
124 §	CALR probe S0999-L26702		19p13.2	L367fs*46=c.1092_1143del52	
130 §	CALR probe S1001-L26517		19p13.2	K385fs*47=c.1154_1155insTTGTC	
136	Reference probe 16316-L25926	3q21.3			
142	Reference probe 07387-L26769	12q13.11			
148	Reference probe 10663-L11245	6p12.2			
154	Reference probe 13781-L15275	11p14.1			
160	Reference probe 17621-L21665	10q22.2			
167 § *	JAK2 probe 16924-L21237		9p24.1	N542_E543del=c.1624_1629delAATGAA	
172 § *	JAK2 probe 16924-L21238		9p24.1	E543_D544del=c.1627_1632delGAAGAT	
175 § JK	MPL probe S1048-SP0405-L21261		1p34.2	W515K=c.1543_1544TG>AA	
182 § JK	MPL probe S1048-SP0405-L20503		1p34.2	W515L=c.1544G>T	
190	Reference probe 11556-L26717	5q31.2			
199 § JK	KIT probe 17722-SP0542-L23707		4q12	D816V=c.2447A>T	
215 § Σ	JAK2 probe 05672-L21576		9p24.1	V617F=c.1849G>T	
230	Reference probe 17130-L26574	11p11.2			
239	Reference probe 05386-L26770	12p11.21			
245	Reference probe 13572-L26953	1q23.2			
265	Reference probe 12434-L26073	14q24.3			
276	Reference probe 16270-L26771	20q11.23			
288	Reference probe 05713-L20268	2p11.2			
297	Reference probe 04570-L20036	16q13			
313	Reference probe 04833-L20693	5p13.2			
328	Reference probe 13397-L26608	6q12			
338	Reference probe 12785-L15496	2q12.3			

§ Mutation-specific probe. This probe will generate a signal when the mutation is present.

JK This probe consists of three parts and has two ligation sites.

* When both probe signals at 167 nt and 172 nt have a similar height to each other it is indicative for JAK2 E543_D544del mutation. When only the probe signal at 167 nt is present, it is indicative for JAK2 N542_E543del mutation.

Σ This probe can have very low unspecific background signal detected also in healthy patient samples. This background signal has never exceeded the signal detected with JAK2 V617F samples with 1% allele burden in the internal quality tests at MRC-Holland. See Figure 2 for further information on this.

Note: Please notify us of any mistakes: info@mlpa.com. The identity of the genes detected by the reference probes is available in Table 2b.

Table 2a. P520-A1 MPN mix 2 probes arranged according to their chromosomal location

Length (nt)	SALSA MLPA probe	Gene / Exon / Mutation details	Ligation site	Partial sequence (24 nt adjacent to ligation site)	MV location (HG18)
MPL gene, at 1p34.2, indicated ligation sites are in NM_005373.2. Mutations in the thrombopoietin receptor (MPL) gene, most commonly W515L and W515K mutations in exon 10, are detected in patients with ET and PMF, which can support diagnosis when JAK2 is not mutated. W515L and W515K are reported to be the predominant MPL mutations in MPNs (Ma W et al. 2011, <i>Diagn Mol Pathol</i> , 20:34-9).					
182 § JK	S1048-SP0405-L20503	MPL, exon 10 W515L=c.1544G>T	1589-1590 and 25nt after ex 10	GCTGCTGATGTT-46nt spanning oligo-TGGCGGGTGAACC	01-043.587.563
175 § JK	S1048-SP0405-L21261	MPL, exon 10 W515K=c.1543_1544TG>AA	1589-1590 and 25nt after ex 10	GCTGCTGAGGAA-46nt spanning oligo-TGGCGGGTGAACC	01-043.587.563
KIT gene, at 4q12, indicated ligation sites are in NM_000222.2. Somatic activating KIT D816V mutation is present in more than 97% of systemic mastocytosis patient samples (Erben P et al. 2014, <i>Ann Hematol</i> , 93:81-8). This D816V mutation results in ligand-independent activation of KIT tyrosine kinase and detection of this point mutation aids in prediction of response to tyrosine kinase inhibitor (TKI) therapy (Imatinib).					
199 § JK	17722-SP0542-L23707	KIT, exon 17 D816V=c.2447A>T	2495-2495 and 2535-2534 reverse	CATTCTTGATGA-38nt spanning oligo-CCATGATGAAGG	04-055.293.998
JAK2 gene, at 9p24.1, indicated ligation sites are in NM_004972.3. Discovery of a JAK2 mutation common to MF, PV and ET has linked these diseases on a molecular level and diagnostic criteria for MPN include detection of a clonal marker e.g. JAK2 V617F mutation in exon 14 or exon 12 mutations. JAK2 is mutated in 97% of all patients with PV, 55% of patients with ET and 65% of patients with PMF (Tefferi A, 2010, <i>Leukemia</i> , 24:1128-38). The most common JAK2 mutations in exon 12 are the following 6-bp deletions: N542-E543del and E543-D544del.					
167 § E	16924-L21237	JAK2, exon 12, N542-E543del =c.1624_1629delAATGAA	2126-2127	AAATCAGAGAT-TTGATATTTGTA	09-005.059.976
172 § E	16924-L21238	JAK2, exon 12, E543-D544del =c.1627_1632delGAAGAT	2126-2127	AAATCAGAAAT-TTGATATTTGTA	09-005.059.976
215 § Σ	05672-L21576	JAK2, exon 14 V617F=c.1849G>T	2343-2342 reverse	GTCTCCACAGAA-ACATACTCCATA	09-005.063.725
CALR gene, at 19p13.2, indicated ligation sites are in NM_004343.3. Majority of patients with ET or PM that are negative for JAK2 and MPL carry a somatic mutation in CALR gene. A 52-bp deletion (type 1=L367fs*46) and 5-bp insertion (type 2=K385fs*47) are the most common mutations found in CALR (53% and 32%, respectively), resulting in a frameshift to an alternative reading frame (Klampff T et al. 2013, <i>N Engl J Med</i> , 369:2379-90; Nangalia J et al. <i>N Engl J Med</i> , 369:2391-405).					
130 §	S1001-L26517	CALR, exon 9 K385fs*47=c.1154_1155insTTGTC	1234-1235	CAGAGGACATTA-TTCGGAGGATGA	19-012.915.595
124 §	S0999-L26702	CALR, exon 9 L367fs*46=c.1092_1143del52	1224-1223 reverse	CTTGCTCTGTC-TCTCTGCTCTGT	19-012.915.528

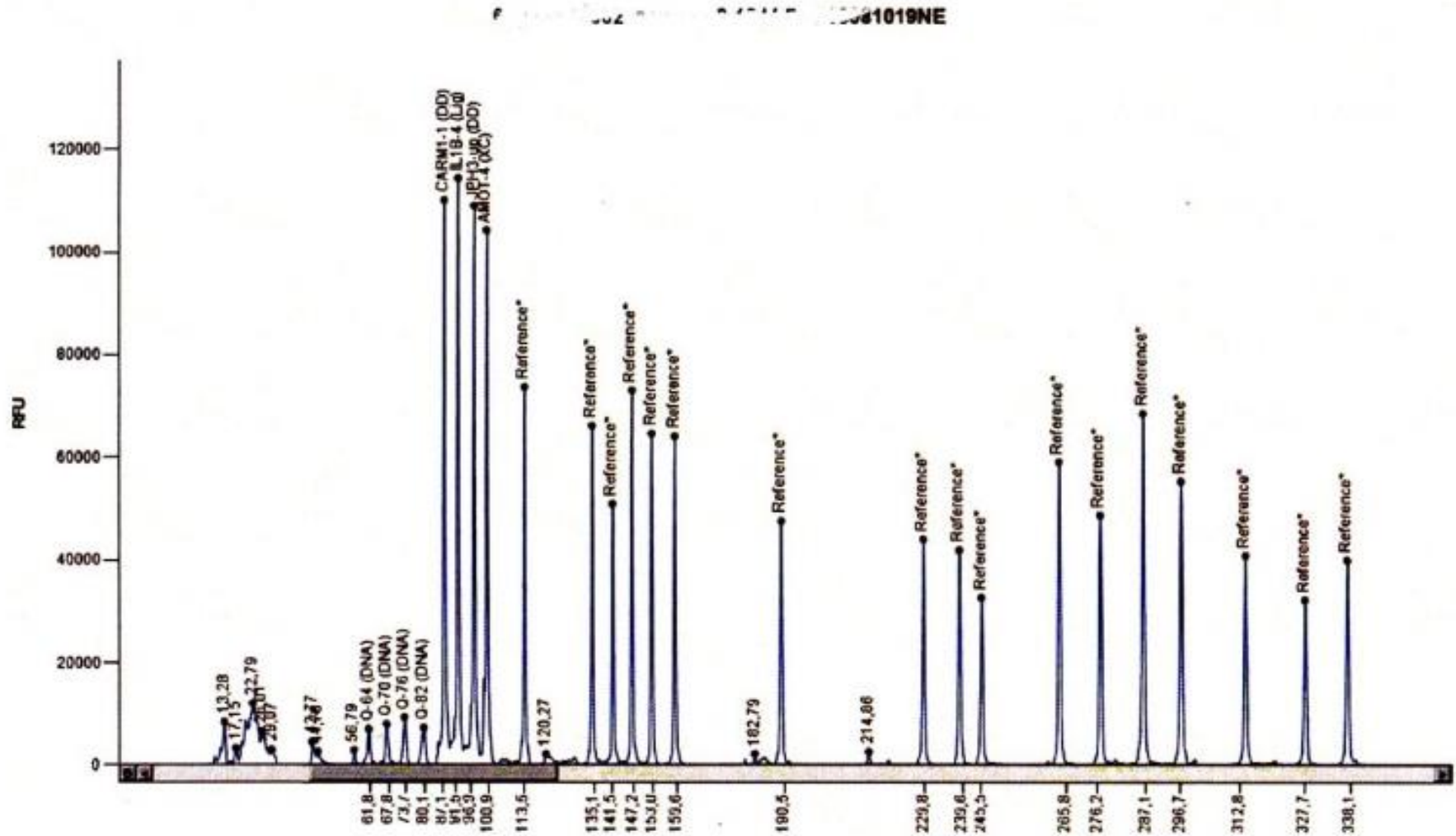
§ Mutation-specific probe. This probe will generate a signal when the mutation is present.

JK This probe consists of three parts and has two ligation sites.

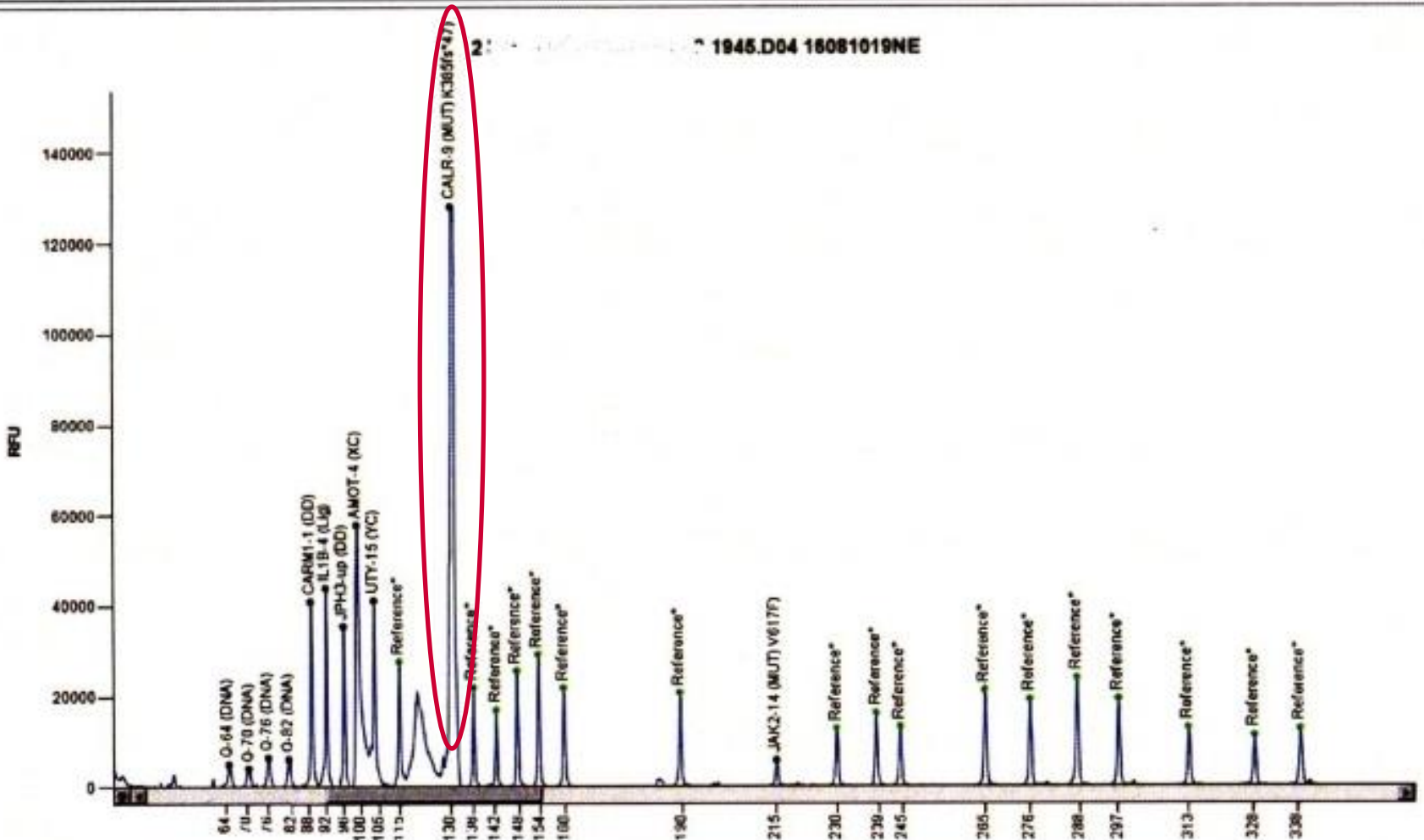
Σ This probe can have very low unspecific background signal detected also in healthy patient samples. This background signal has never exceeded the signal detected with JAK2 V617F samples with 1% allele burden in the internal quality tests at MRC-Holland. See Figure 2 for further information on this.

E When both probe signals at 167 nt and 172 nt have a similar height to each other it is indicative for JAK2 E543_D544del mutation. When only the probe signal at 167 nt is present, it is indicative for JAK2 N542_E543del mutation.

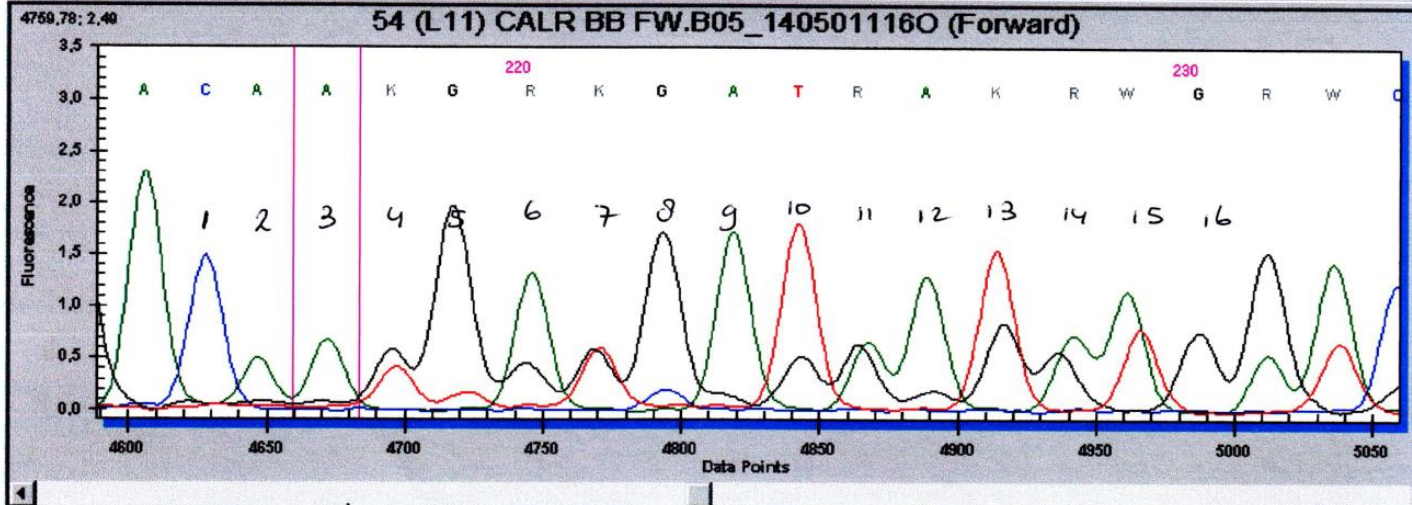
Note: Exon numbering may differ from literature! Complete probe sequences are available on request: info@mlpa.com. Please notify us of any mistakes: info@mlpa.com.



MLPA CALR mut type 2



Base Number	310				320				330				340			
Codon Number	102	103	104	105	106	107	108	109	110	111	112	113	114	115		
Reference AA Translation	E	D	K	E	D	D	E	D	K	D	E	D	E	F		
CALR BB FW+ CALR AA Rev.txt	CA	GAG	GAC	AAG	GAG	GAT	GAT	GAG	GAC	AAA	G-AT	GAG	GAT	C		
54 (L11) CALR BB FW.B05_140501	CA	GAG	GAC	AAK	GRK	GAT	RAK	RWG	RWC	AAR	GAMT	RAG	RWT	F		
Differences				?	??		? ?	??	??		? <+?	?	??	?		
Consensus	CA	GAG	GAC	AAK	GRK	GAT	RAK	RWG	RWC	AAR	GAMT	RAG	RWT	F		
Consensus AA Translation		E	D	KN	*	D	*	*	*	K	DE	X	*	X		



L11 = C. 1154_1155 ins TTGTC

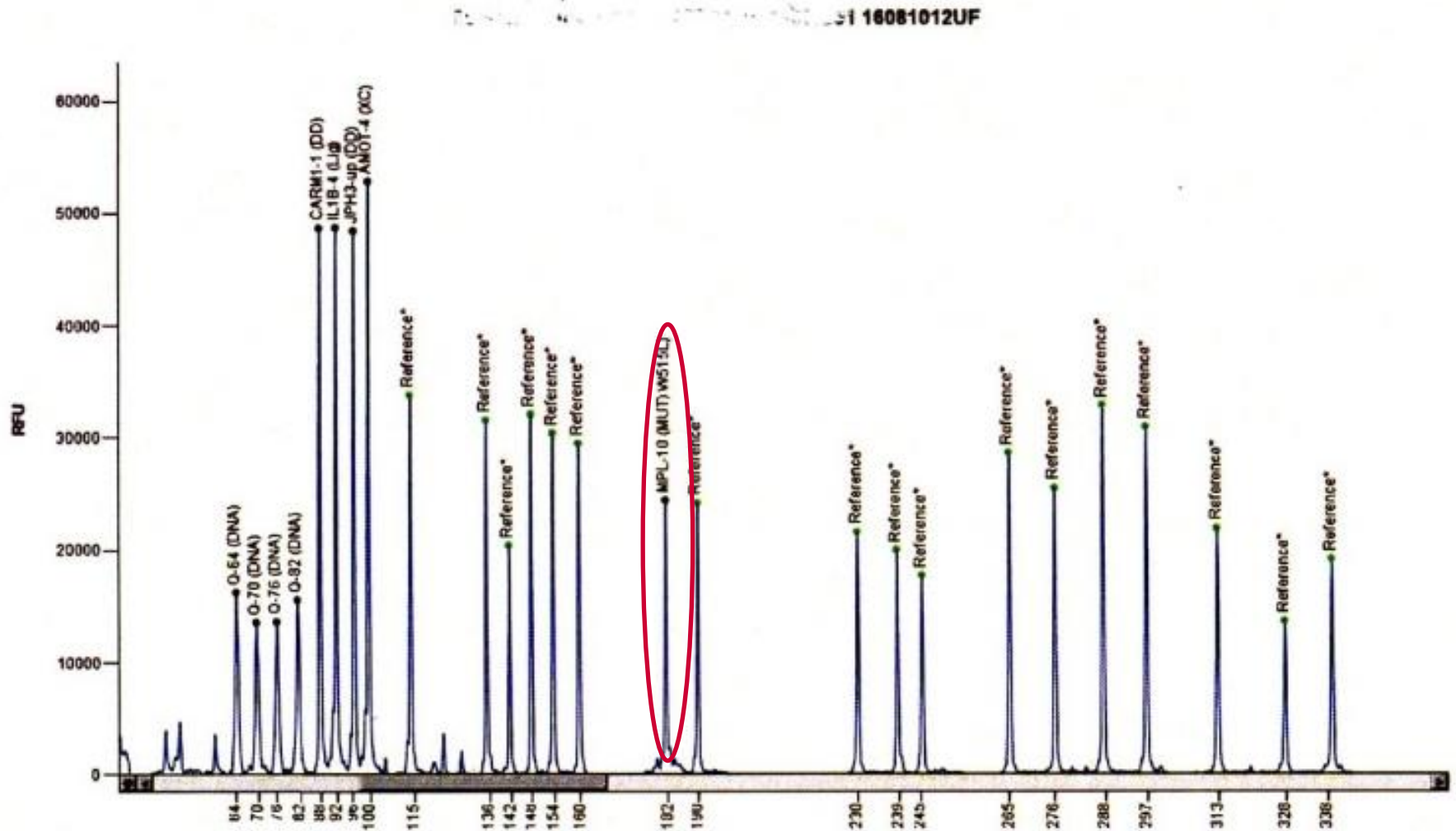
1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16

C A A G G A G G A T G A T G A G

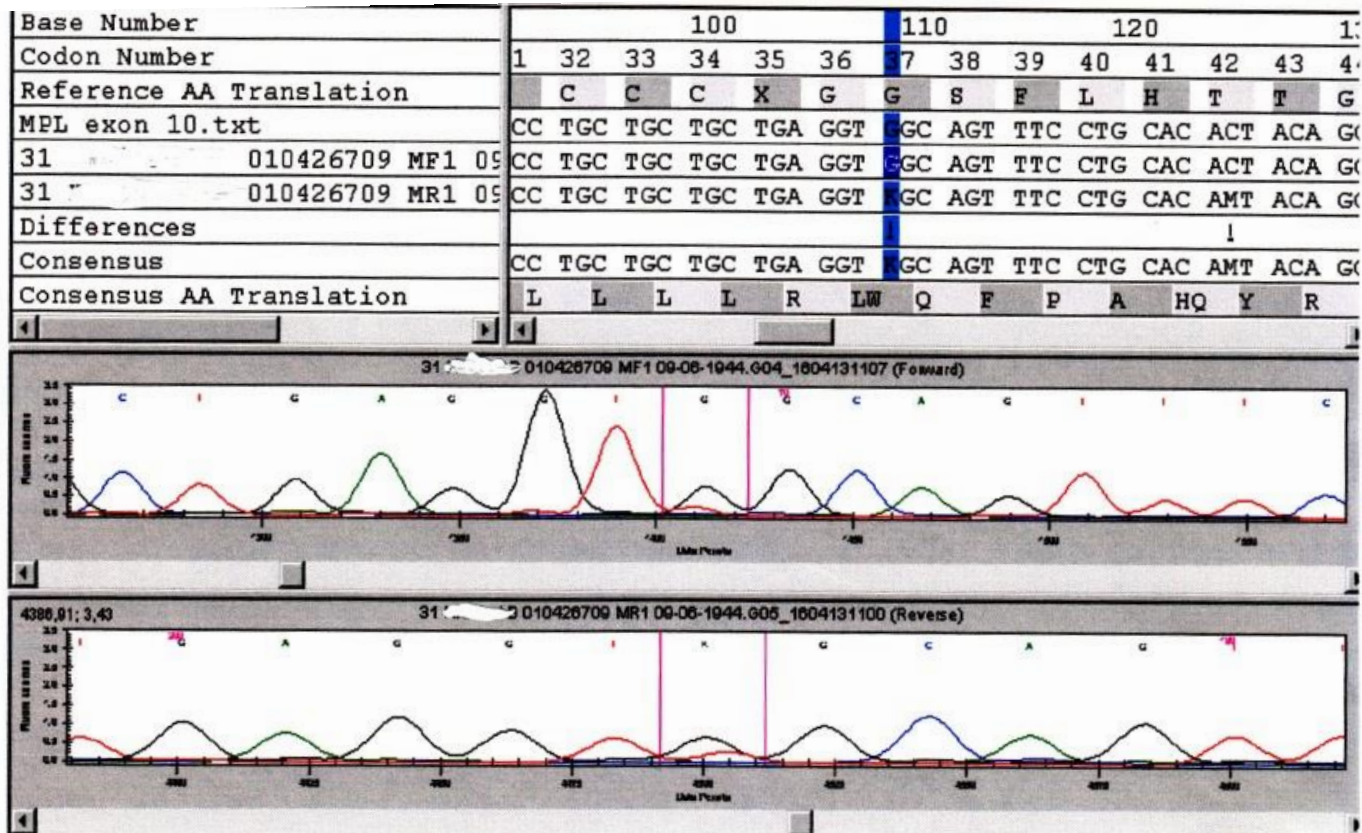
C A A TTGTC G G A G G A T G

↓
inserie.

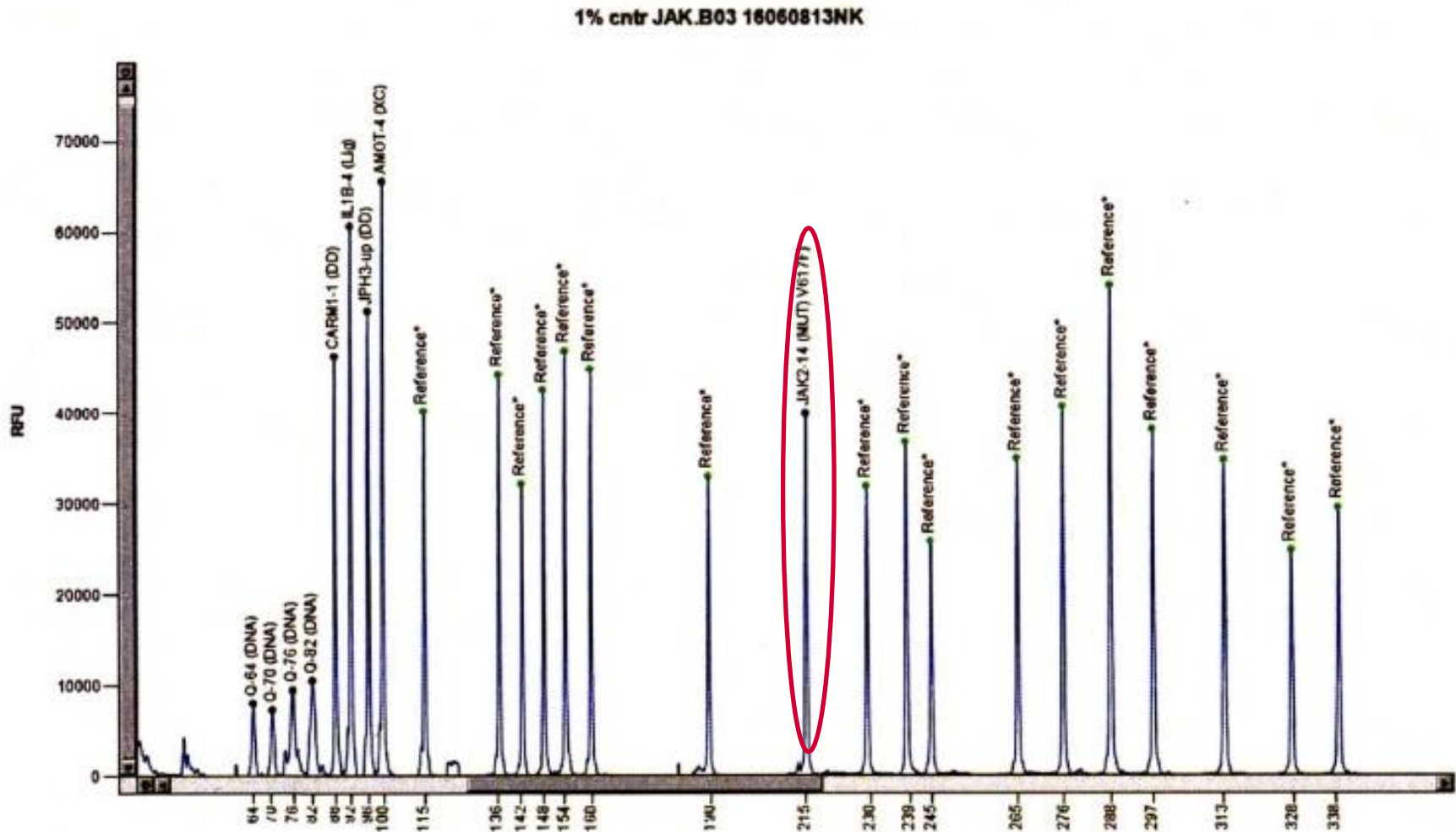
MLPA MPL mutatie



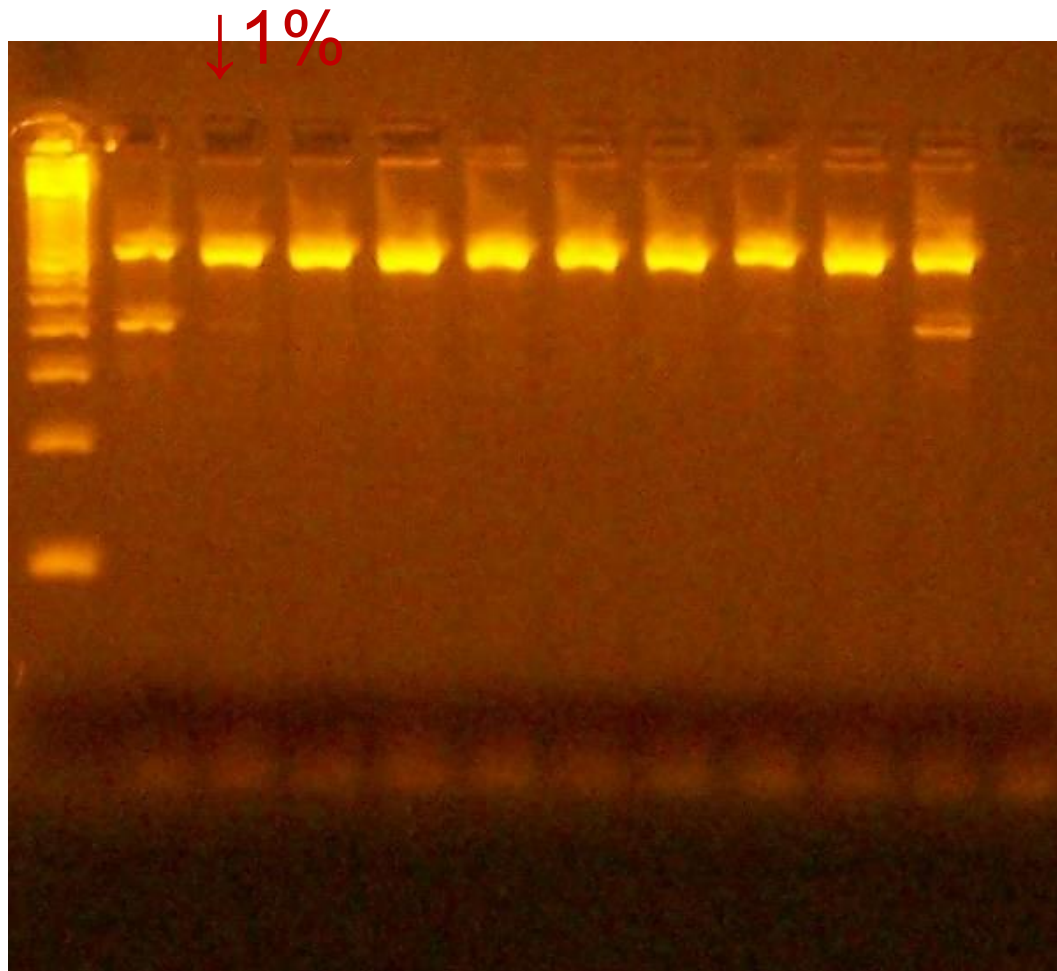
Sequentie analyse MPL



MLPA JAK V617F 1% load



PCR JAK-2 V617F



PAL: nieuwe locatie DNA

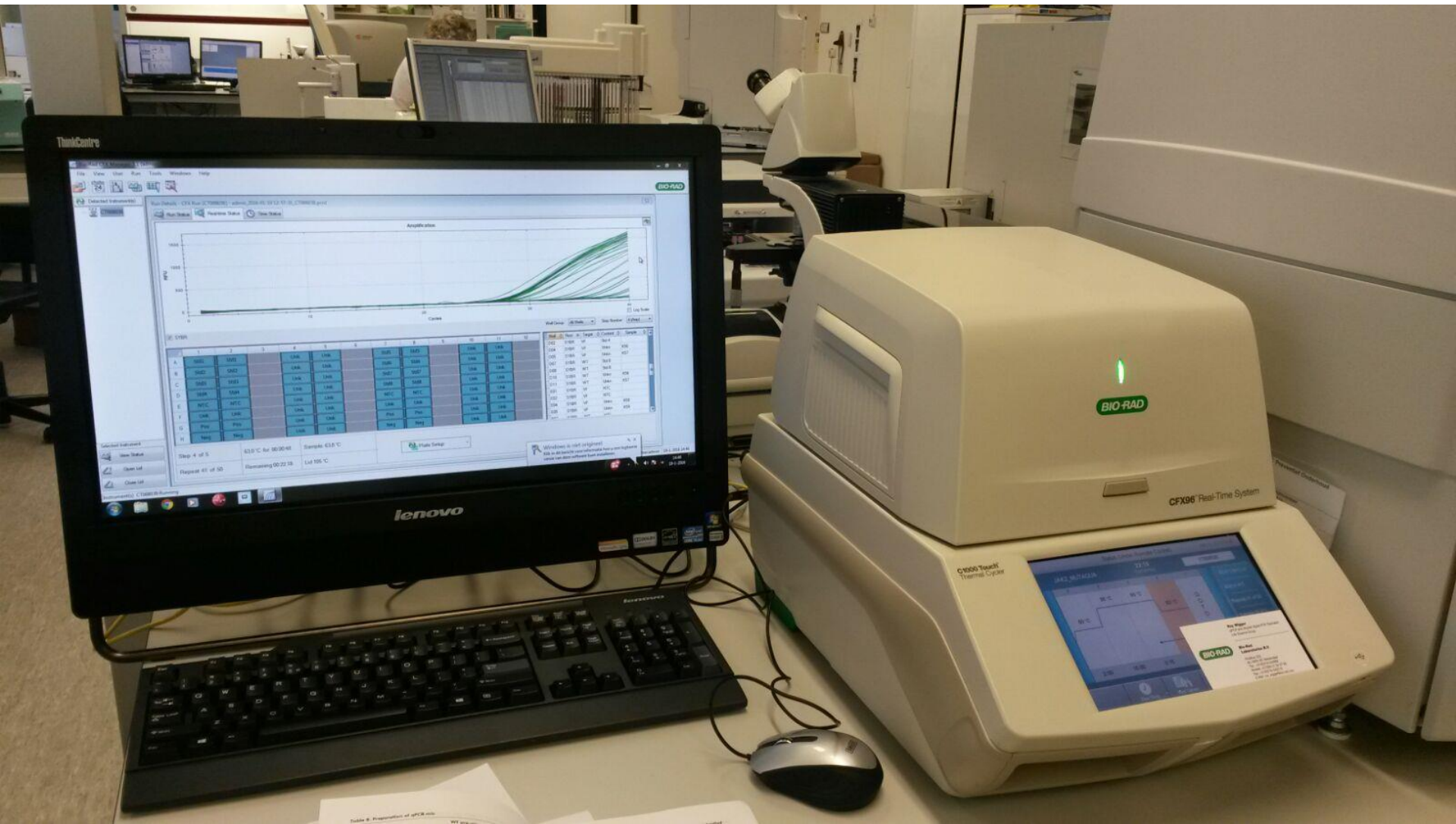








Nieuw realtime PCR apparaat.



Casus Marjan



Hematologie/serologie

Hemoglobine	H	12.5	mmol/L	(8.5 - 11.0)
Hematocriet	H	0.59	L/L	(0.41 - 0.51)
Erytrocyten		5.93	10e12/L	(4.60 - 6.20)
MCV		100	fL	(80 - 100)
RDW		14.5	%	(11.0 - 16.0)
vitamine B12		692	pmol/L	(130 - 700)
foliumzuur		7	nmol/L	(5 -)
Ferritine		38	ug/L	(25 - 250)
IJzer		26.3	umol/L	(14 - 28)
Transferrine		2.98	g/L	(2.00 - 3.60)
TIJBC		75	umol/L	(45 - 80)
Verzadigingspercentage		35	%	(20 - 60)
Trombocyten		277	10e9/L	(150 - 400)
Leukocyten		6.2	10e9/L	(4.3 - 10.0)

Machine diff

Basofielen		< 0.1	10e9/L	(0.0 - 0.15)
Eosinofielen		0.2	10e9/L	(- 0.7)
Lymfocyten		1.2	10e9/L	(1.0 - 4.0)
Neutrofielen		4.2	10e9/L	(1.8 - 7.3)
Monocyten		0.6	10e9/L	(0.2 - 1.0)

JAK2 V617F

Zie Opm. → neg

Zie uitslagbrief.

Klinische Chemie algemeen

Creatinine		78	umol/L	(59 - 104)
MDRD (e-GFR)		> 60	mL/min	(60 -)
Urinezuur	H	0.48	mmol/L	(0.20 - 0.42)
Natrium		139	mmol/L	(135 - 145)
Kalium		3.7	mmol/L	(3.5 - 5.0)
Calcium		2.37	mmol/L	(2.20 - 2.65)
Albumine		42	g/L	(35 - 50)
Bilirubine	H	24	umol/L	(- 17)
Gamma GT		18	E/L	(- 50)
Alkalische fosfatase		100	E/L	(30 - 100)
ASAT		36	E/L	(- 37)
LDH		380	E/L	(- 450)
Testosteron		10.9	nmol/L	(5.0 - 16.5)
Vrij testosteron index		32		(20 - 96)
DHEAS		3.9	umol/L	(0.0 - 9.9)
SHBG		34	nmol/L	(10 - 57)

Eiwitten en Markers

CRP		< 5	mg/L	(0 - 10)
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Verzendbepalingen

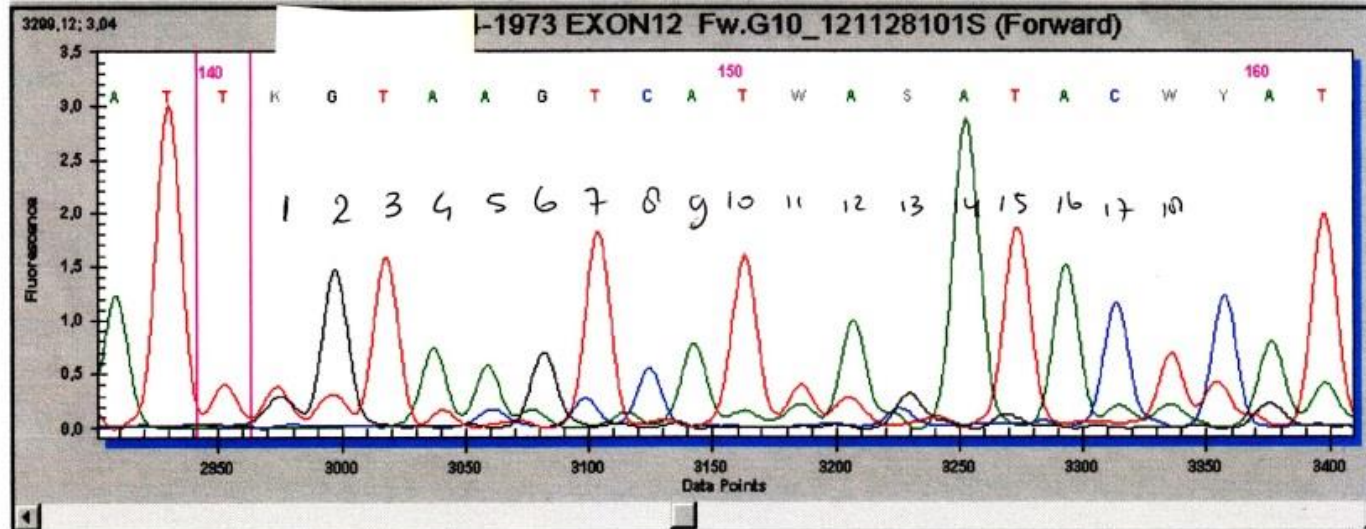
Erytropoetine	L	<1	IU/L	(4.0 - 29.0)
---------------	---	----	------	----------------

Man

69 Jaar

Wat valt op?

Base Number	190			200			210			220			
Codon Number	64	65	66	67	68	69	70	71	72	73	74	75	76
Reference AA Translation	L	I	F	V	S	H	X	I	L	I	T	V	F
JAK 2 ex12.txt	TTG	ATA	TTT	GTA	AGT	CAT	TAG	ATA	CTC	ATT	ACT	GTC	TT-?
XON12 Fv	TTG	ATA	TTK	GTA	AGT	CAT	WAS	ATA	CWY	ATW	AMT	GTS	WTAT
Differences			?				?	?	??	<?	?	?	?
Consensus	TTG	ATA	TTK	GTA	AGT	CAT	WAS	ATA	CWY	ATW	AMT	GTS	WTAT
Consensus AA Translation	L	I	FL	V	S	H	*	I	HL	I	NT	V	IL



1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17

Ⓡ T G T A A G T C A T T A G A T A

Ⓟ G T T T C A C A A A A T C A G A

Sequentie met forward primer

TGTTTCACAA AATCAGAAAT GAAGATTGA TATTGTTTCACAA AATCAGAAAT GAAGATTGA
TGTTTCACAA AATCAGAAAT GAAGATTGA TATTGTAAAGTCA TTAGATA

TATTGTAAAGTCA TTAGATA

Sequentie met reverse primer

TGTTTCACAA AATCAGAAAT GAAGATTGA TATTGTTTCACAA AATCAGAAAT GAAGATTGA
ATGAACCAAA TGGTGTTCACAA AATCAGAAAT GAAGATTGA

TATTGTAAAGTCA TTAGATA

TATTGTAAAGTCA TTAGATA

Patiënt

TG TTT CAC AAA ATC AGA AAT GAA GAT TTG ATA TTG TTT CAC AAA ATC AGA
F H K I R N E D L I L F H K I R

AAT GAA GAT TTG ATA TTT GTAAGTCATTAGATA

N E D L I F intron

Wild type

TG TTT CAC AAA ATC AGA AAT GAA GAT TTG ATA TT|T GTAAGTCATTAGATA
F H K I R N E D L I F intron

Insertie (duplicatie) bij patiënt is onderstreept. Plaats van insertie tov wild type is aangegeven met verticale streep in wild type sequentie.

Beschreven in de literatuur als:

F537-I546dup10, F547L mutatie (eiwit)
1608-1640dup133 (DNA)

Originele referentie:

Pietra D, Li S, Brisci A, Passamonti F, Rumi E, Theocharides A, et al.

Somatic mutations of JAK2 exon 12 in patients with JAK2 (V617F)-negative myeloproliferative disorders. Blood 2008;111:1686-9.

Casus 2: vrouw uit 1964

Hb: 7.0
WBC: 12.1
Trombo's: 236
Reti's: 1.2%

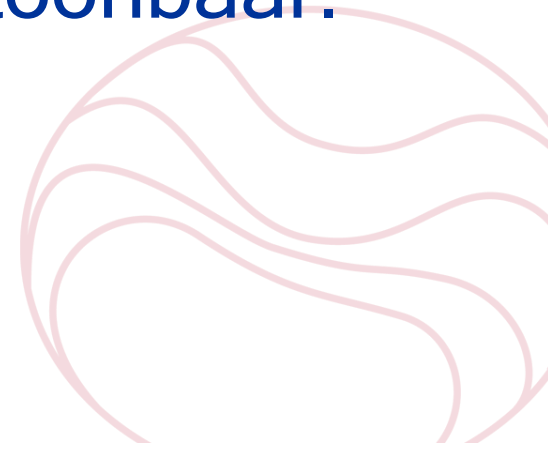
♦ DIFF.	.			
♦ eo's	1		%	0 - 6
♦ baso's	1		%	0 - 2
♦ staven	9	+	%	0 - 4
♦ segmenten	26	-	%	40 - 80
♦ lymfo's	38		%	12 - 46
♦ mono's	8		%	2 - 12
♦ blasten	4	+	%	0 - 0
♦ promyelo's	3	+	%	0 - 0
♦ myelo's	2	+	%	0 - 0
♦ metamyelo	8	+	%	0 - 0
♦ anisocyt.	++			
♦ microcyt.	++			
♦ anisochr.	+			
♦ hypochr.	+++			
♦ bas.punkt.	+			
♦ elliptocyt	+			
♦ polychr.	+			
♦ traandr.cel	+			
♦ erytroblas	3		/100 wbc	
♦ Diff. opmerk	TEKST advies: consult hematoloog (hs)			
♦ Trombocyten	289		10exp 9/l	150 - 400
♦ macro trom.	+			
-----KLINISCHE CHEMIE-----				
♦ Kreatinine	64		umol/l	45 - 90
♦ ASAT	29		u/l	<31
♦ ALAT	25		u/l	<34
♦ LD	914	+	u/l	<250
♦ Alk.fosfat.	109		u/l	40 - 125
♦ Albumine	39		g/l	35 - 52
♦ eGFR CKD-EPI	82		ml/mn/1.73m2	>60
-----KLINISCHE CHEMIE SPECIEEL-----				
♦ CRP	<2.9		mg/l	<10
-----KLIN.CHEM.SPECIEEL, VITAMINE'S---				
♦ Vitamine B12	869	+	pmol/l	130 - 700
♦ 25-OH Vit.D3	10	-	nmol/l	50 - 150

Moleculaire diagnostiek

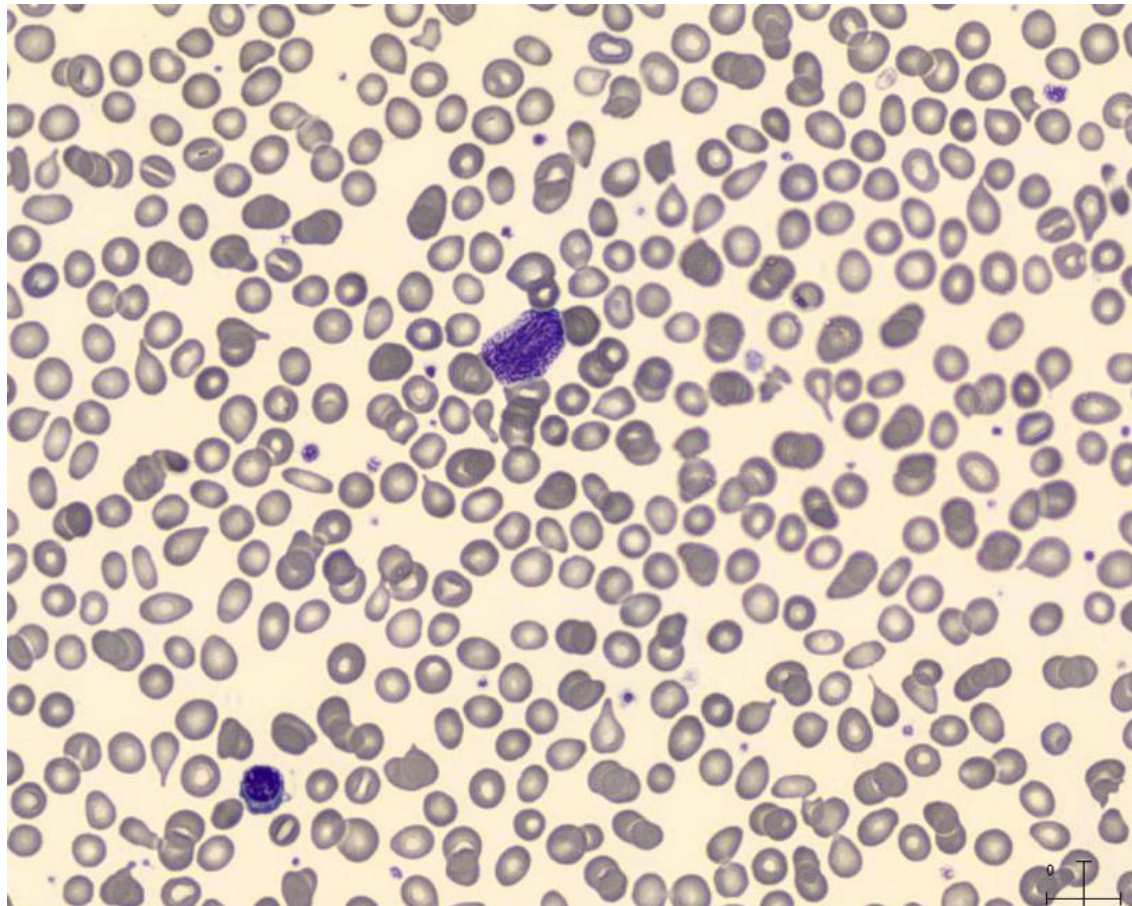
JAK2 V617F: niet aantoonbaar.

MPL W515K/L: niet aantoonbaar.

CALR exon 9: type 1 mutatie aantoonbaar.



Perifeer bloed



Beenmergonderzoek

Flowcytometrie: **Perifeer bloed: 3% myeloide blasten.**
Beenmerg gelijk aan perifeer bloed. (?)

Patholoog (biopt): **Myelofibrose graad 3. Geen blasten toename.**

Klinische chemie:

Conclusie:

Perifeer bloed, waarin een leuko-erytoblastair bloedbeeld met traandruppelcellen en morfologisch circa 2% blasten. Beenmergpreparaten van slechte kwaliteit (dry tap?), waarin een van de beschikbare preparaten een zeer forse toename van morfologisch afwijkende megakaryocyten. Beeld allereerst sterk verdacht voor een myeloproliferatieve aandoening: in casu (primaire) myelofibrose.

Table 7. WHO criteria for overt PMF

WHO overt PMF criteria

Major criteria

1. Presence of megakaryocytic proliferation and atypia, accompanied by either reticulin and/or collagen fibrosis grades 2 or 3*
2. Not meeting WHO criteria for ET, PV, *BCR-ABL1*⁺ CML, myelodysplastic syndromes, or other myeloid neoplasms
3. Presence of *JAK2*, *CALR*, or *MPL* mutation or in the absence of these mutations, presence of another clonal marker,† or absence of reactive myelofibrosis‡

Minor criteria

Presence of at least 1 of the following, confirmed in 2 consecutive determinations:

- a. Anemia not attributed to a comorbid condition
- b. Leukocytosis $\geq 11 \times 10^9/L$
- c. Palpable splenomegaly
- d. LDH increased to above upper normal limit of institutional reference range
- e. Leukoerythroblastosis

Diagnosis of overt PMF requires meeting all 3 major criteria, and at least 1 minor criterion

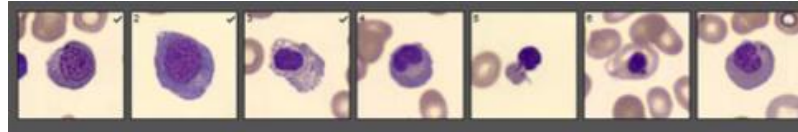
*See Table 8.

†In the absence of any of the 3 major clonal mutations, the search for the most frequent accompanying mutations (eg, *ASXL1*, *EZH2*, *TET2*, *IDH1/IDH2*, *SRSF2*, *SF3B1*) are of help in determining the clonal nature of the disease.

‡BM fibrosis secondary to infection, autoimmune disorder, or other chronic inflammatory conditions, hairy cell leukemia or other lymphoid neoplasm, metastatic malignancy, or toxic (chronic) myelopathies.

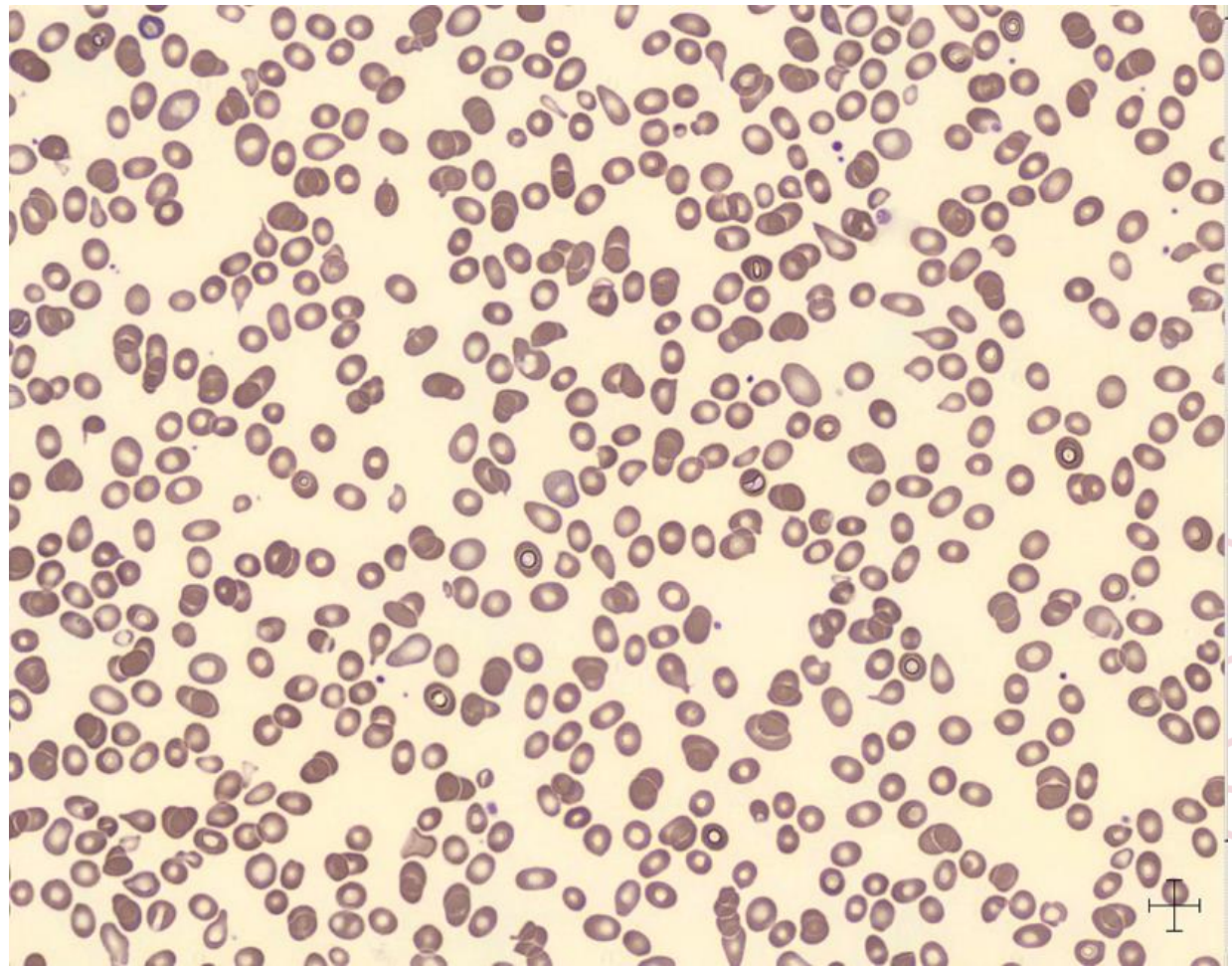


Casus 3: vrouw uit 1964



Hb: 3.2
WBC: 5.5
Trombo's: 90
Reti's: 2.6%

♦ DIFF.	.
♦ eo's	0
♦ baso's	1
♦ staven	1
♦ segmenten	56
♦ lyfmo's	29
♦ mono's	7
♦ blasten	2
♦ promyelo's	1
♦ myelo's	2
♦ metamyelo	1
♦ poikilocyt	+
♦ fragmento	+
♦ traandr.cel	++
♦ erythroblas	4
♦ Trombocyten	90
♦ thr.aggr.	afw



Moleculaire diagnostiek

JAK2 V617F: niet aantoonbaar.

MPL W515K/L: niet aantoonbaar.

CALR exon 9: niet aantoonbaar.



Casus 3: vrouw uit 1964

LD	>4000	++	u/l	<250
Bili. totaal	31	+	umol/l	<17
Bili. direct	4		umol/l	<5
Alk.fosfat.	59		u/l	40 - 125
GGT	17		u/l	<38
Albumine	38		g/l	35 - 52
eGFR CKD-EPI	>90		ml/mn/1.73m2	>60
Spijtserum	g8a			
-----KLINISCHE CHEMIE SPECIEEL-----				
IJzer	49	+	umol/l	9 - 30
Transferrine	2.54		g/l	2.0 - 3.6
Transf.verz.	77	+	%	<45
Ferritine	480	+	ug/l	20 - 150
Haptoglobine	<0.08	--	g/l	0.3 - 2.0
-----KLIN.CHEM.SPECIEEL, VITAMINE'S---				
Vitamine B12	44	--	pmol/l	130 - 700
Foliumzuur	13		nmol/l	5 - 36

Beenmergonderzoek

Flowcytometrie:

Perifeer bloed met een myeloïde blasten populatie met een omvang van circa 1% van de leukocyten. Beenmerg?

Patholoog (biopt): met

cristabiopt: representatief biopt met zeer celrijk merg (100%) enige dysmegakaryopoïese en hyperplasie van de erythroïde reeks met forse linksverschuiving. Geen significante blastentoeename. Geen fibrose. Beeld passend i.h.k.v. vitamine B12 deficiëntie. Geen myeloproliferatieve neoplasie of overtuigende aanwijzingen voor MDS.

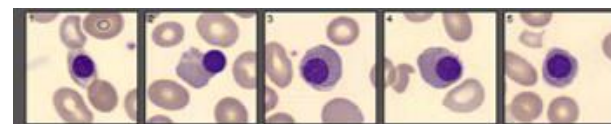
Klinische chemie:

Conclusie:

Perifeer bloed met opvallende vormafwijkingen van de rode bloedcellen. Beenmerg met een verhoogd aantal megakaryocyten en actieve erythropoïese. Vooralsnog past dit bij de ernstige vitamine B12 deficiëntie. Een (prefibrotisch stadium van) myelofibrose lijkt op basis van de morfologie minder waarschijnlijk op dit moment gezien de o.a. relatief normale morfologie van de megakaryocyten. Graag deze uitslag interpreteren in samenhang met de PA.

Casus 3: vrouw uit 1964

5 dagen na B12 injecties



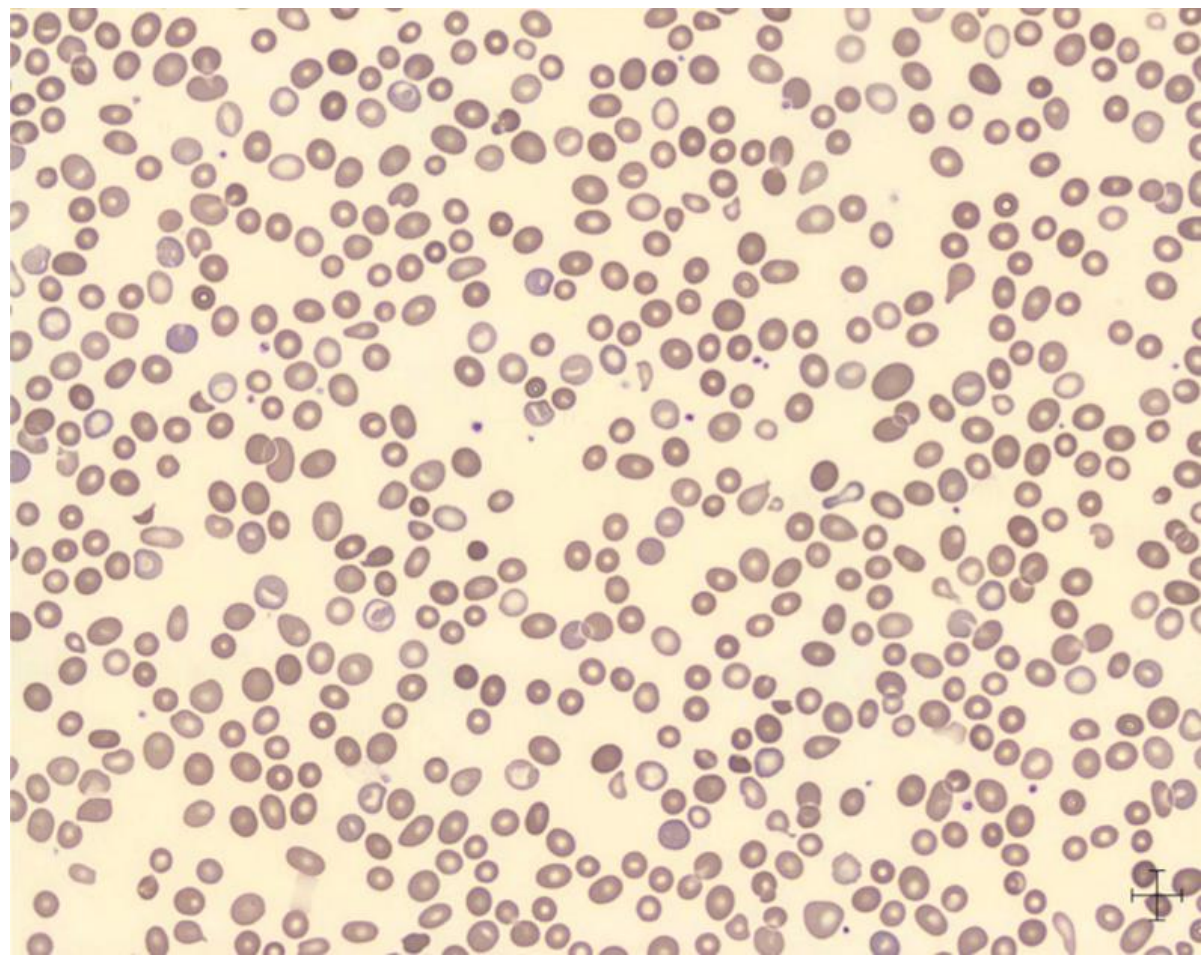
Hb: 5.7

WBC: 4.5

Trombo's: 117

Reti's: 22%

❖ DIFF.	.
❖ eo's	2
❖ baso's	0
❖ staven	1
❖ segmenten	65
❖ lymfo's	25
❖ mono's	7
❖ bas.punkt.	+
❖ poikilocyt	++
❖ traandr.cel	+
❖ erytroblas	3
❖ Trombocyten	117
❖ macro trom.	+



Casus 3: vrouw uit 1964

Bepaling	Eenheid	Ref. waarde	12-05-2016 08:58 S. Kuipers	12-05-2016 11:24 S. Kuipers	17-05-2016 11:33 S. Kuipers	17-05-2016 11:34 S. Kuipers	31-05-2016 09:52 S. Kuipers
HEMATOLOGIE							
Hemoglobine	mmol/l	7.5 - 10.0	3.2 L		5.7 L		7.3 L
Hematocriet	l/l	0.35 - 0.45	0.15 L		0.29 L		0.37
Erytrocyten	10exp12/l	4.00 - 5.00	1.29 L		2.66 L		3.78 L
MCV	fl	80 - 100	116 H		108 H		98
MCHC	mmol/l	20 - 23	21		20		20
MCH	amol	1750 - 2100	2460 H		2140 H		1930
RDW		12.1 - 14.1	20.8 H		23.9 H		18.9 H
Reticulocyt	%	0.5 - 2.5	2.6 H		22.4 H		2.6 H
Leukocyten	10exp 9/l	4.0 - 10.0	5.5		4.5		8.7
Neutrof.tot	10exp 9/l	1.8 - 7.3	3.32		2.89		5.64
Eosinofielen	10exp 9/l	0 - 0.7					0.26
DIFF.							
eo's	%	0 - 6	0		2		3
baso's	%	0 - 2	1		0		1
staven	%	0 - 4	1		1		
segmenten	%	40 - 80	56		65		65
lymfo's	%	12 - 46	29		25		23
mono's	%	2 - 12	7		7		8
blasten	%	0 - 0	2 H				
promyelo's	%	0 - 0	1 H				
myelo's	%	0 - 0	2 H				
metamyelo	%	0 - 0	1 H				
poikilocyt			+		++		
fragmento			+				
traandr.cel			++		+		+
erythroblas	/100 wbc		4		3		
bas.punkt.					+		
Trombocyten	10exp 9/l	150 - 400	90 L		117 L		489 H
thr.aggr.			afw				
macro trom.					+		
Bezinking	mm/uur	<30	9				
BLOEDGROEPENSEROLOGIE							
Bloedgr/Rh			A pos	A pos			
Kruisbloed			<Memo>				
IrregulairAS			neg	neg			
Dir. Coombs			neg				
STOLLING,TROMBOSE,FIBRINOLYSE							
PT (sec)	sec	9 - 14	11				
APTT	sec	ca.35	21				
BEENMERG							
Beenmerg	nr.	.				klaar	
KLINISCHE CHEMIE							
Natrium	mmol/l	135 - 145	139		144		137
Kalium	mmol/l	3.5 - 5.0	3.3 L		4.1		4.3
Calcium	mmol/l	2.10 - 2.55	2.21		2.24		2.33
Kreatinine	umol/l	45 - 90	41 L		35 L		42 L
eGFR CKD-EPI	ml/mn/1.73	>60	>90		>90		>90
ASAT	u/l	<31	105 H				
ALAT	u/l	<34	39 H				
LD	u/l	<250	>4000 H		2072 H		284 H

Huisarts belt over labuitslagen van man uit 1968.

Bezinking	2	mm/uur	(- 15)
Hemoglobine	9.9	mmol/L	(8.5 - 11.0)
Hematocriet	0.49	L/L	(0.41 - 0.51)
vitamine B12	242	pmol/L	(130 - 700)
Trombocyten	H 813	10e9/L	(150 - 400)
Leukocyten	H 10.7	10e9/L	(4.3 - 10.0)
Machine diff			
Basofielen	0.1	10e9/L	(0.0 - 0.15)
Eosinofielen	0.4	10e9/L	(- 0.7)
Lymfocyten	2.5	10e9/L	(1.0 - 4.0)
Neutrofielen	7.1	10e9/L	(1.8 - 7.3)
Monocyten	0.5	10e9/L	(0.2 - 1.0)
Creatinine	64	umol/L	(59 - 104)
MDRD (e-GFR)	> 60	mL/min	(60 -)
Gamma GT	7	E/L	(- 50)
Alkalische fosfatase	68	E/L	(30 - 100)
ALAT	16	E/L	(- 41)
Glucose nuchter plasma	5.2	mmol/L	(4.0 - 6.1)
TSH screening			
TSH	2.8	mE/L	(0.4 - 4.0)
Vitamine D 25-OH	L 38	nmol/L	(50 - 200)
Prostaat Specif. Ag	1.6	ug/L	(0.0 - 4.0)
***** < EINDE > *****			

Wat vinden we hiervan?
Wat gaan we doen?

